



Anthracnose Disease of Landscape Plants¹

Stephen H. Brown, Philip F. Harmon, Sheryl van der Heiden, Ryan Czaplewski, Ralph Mitchell, and David Outerbridge²

Target Audience

This publication is intended to provide information to professional landscapers, pest control operators, Extension agents, and homeowners on how to recognize and potentially manage anthracnose diseases of landscape plants.

Introduction

Fungi make up approximately 90% of known plant pathogens. One of the most common fungal diseases of plants is anthracnose. The name anthracnose represents a group of related fungal diseases that affect many different landscape and crop plants. These anthracnose diseases produce similar sets of symptoms with a few consistent characteristics. Though members of several genera are known to cause anthracnose, the most causative agents belong to the *Colletotrichum* genus. In Florida, *C. gloeosporioides* is the most common species causing anthracnose in the landscape. The disease can be a significant problem during stressful, hot, and humid conditions.

Life Cycle

The *Colletotrichum* species that cause anthracnose are common in the Florida landscape. When not causing disease, they help decompose dead or damaged plant tissues, acting as saprophytes (living on dead plants). These fungi produce spores that are sticky and not easily spread through the air. Instead, water splashing from rainfall and irrigation more commonly transports the spores to their host. They may spread from diseased or decaying plant tissue to healthy plants from people's hands or tools. If the fungal spores (Figure 1) settle on the surface of a host plant, the right level of moisture allows them to produce short hyphae, which terminates in anchoring and penetrating structures called appressoria. The appressoria produce penetration pegs that work their way through the cuticle of living plants into the epidermis and deeper plant tissues. For a time, the invading hyphae may feed on living cells without damaging them (biotrophy). The young hyphae may also go into quiescence, a dormant period in the fungal life cycle that can extend from days to months. Therefore, in a biotrophic or quiescent stage, the pathogen

does not produce any noticeable symptoms. Its presence may be confirmed with molecular lab analyses.

Physical damage, chemical burns, or stress from notable environmental changes in temperature or moisture can initiate the active phase of the disease cycle. Extended periods of wet and humid weather in the form of rain, dew, or fog produce environmental conditions that favor fungal growth and increase the plant's susceptibility to disease. When leaf surfaces remain wet for four or more hours after an irrigation event or rainstorm, the anchored fungus begins to kill plant tissues and feed from the resulting dead (necrotic) plant organs. Even moisture from dew can extend the time the plant remains wet, increasing its chances to infiltrate plant tissues. Although a wound on an infected plant is not necessary for the pathogen to enter its host, physical damage can increase how severe the disease symptoms may become. As lesions form on the diseased plant parts, the pathogen continues to produce more of the sticky spores that continue the cycle of infection if favorable conditions persist.

Symptoms and Signs

Anthracnose can cause symptoms on a diverse range of plant parts, including leaves, stems, fruits, and flowers. Anthracnose causes death and discoloration of plant tissues and may be confused with numerous other plant diseases or damages. Salt burn from excess salinity can produce anthracnose-like symptoms, such as tip and marginal leaf burn, leaf browning, and defoliation. Sunscald may be mistaken for anthracnose as well. It is important to keep in mind that, in many cases, symptoms can have multiple causes simultaneously (abiotic damage and multiple diseases on the same plant parts).

Anthracnose symptoms vary depending on the fungal species complex, the host plants, and weather conditions. The following list describes a range of typical symptoms of the disease, usually occurring in various combinations:

- Brown-to-black spots appear on leaves surrounded by yellow halos and concentric rings.
- Irregular yellow or brown spots on leaves darken and merge as they age, sometimes covering the leaves.

- Leaves and tender shoots are blighted by lesions and withering.
- Chlorotic-to-necrotic leaf burn starts with a bright yellow color on the margins. As the chlorosis progresses toward the center and lower end of the leaf, the marginal leaf tissue turns brown and becomes distorted.
- Marginal leaf burn and leaf spots occur simultaneously.
- Large areas of dead tissue cause cupping and distortion of expanded leaves.
- Twigs and branches die back.
- Leaf loss occurs in severe cases and seasonally. Leaves regenerate with new growth.
- Small, dark, sunken lesions of irregular shape appear on fruit and spread as the disease progresses, degrading the quality of maturing fruit, which eventually rots on the tree or in storage.

Anthracnose rarely causes severe and lasting damage to plants. For some plants, the infection is a seasonal occurrence and part of the replacement of old leaves with new ones. This commonly occurs with queen crape myrtle (*Lagerstroemia speciosa*, Figure 45) and many of the trumpet trees (*Handroanthus* spp.; *Tabebuia* spp., Figure 44). Other plants, most noticeably the corn plant (*Draceana fragrans*, Figure 20), seem unable to rid themselves of the disease because of persistent and unfavorable environmental conditions.

Susceptibility and severity of anthracnose infections may increase when plants, poorly adapted to wet conditions, are growing in moisture-retaining areas in the landscape. However, anthracnose-causing fungal species may be part of the microflora of landscape plants. Changes in environmental conditions, the onset of leaf senescence, nutrient deficiencies, or a decline in the plant's overall health may facilitate the fungi's transition from commensal or mutualistic symbiont (not harmful or even beneficial to the plant host) to opportunistic pathogen (causing disease because of the decline in plant health).

Signs of the anthracnose pathogen are more difficult to see than the symptoms on the plant. These include structures and spores produced by the pathogen on symptomatic plant tissues. The most distinct anthracnose signs are hairlike structures called setae (singular: seta) (Figure 2). Visible with a magnifying glass, setae are distinctly black in color and arranged in small circular groups of protruding spore-producing structures called acervuli. Setae are not always present. They may be cultivated and observed by enclosing symptomatic plant material in a resealable bag with a moist paper towel for 24 hours (incubation).

Masses of sticky spores may also be visible after incubation or extended periods of wet conditions. *Colletotrichum* spores frequently form orange-to-salmon-colored clusters and can form tendrils that exude from blister-like

structures embedded in the host plant's dead tissue (Figure 3).

Management

These diseases are often effectively controlled by following good sanitary and cultural practices and are rarely serious enough to warrant chemical control. The following are ways to manage the disease:

Manual

- Increase air circulation by removing some plants or spacing them further apart.
- For herbaceous perennial and palms, remove and discard diseased leaves depending on species tolerance.
- For woody perennials and trees, prune out infected stems and branches about four inches below diseased areas, using a sanitizer on pruning tools between cuts.
- Rake and remove fallen leaves.
- Increase sun exposure or decrease shade.
- Decrease volume or duration of irrigation depending on species recommendations.
- Restrict irrigation or watering to morning hours after dewfall to reduce the duration the plant is wet.
- Install drip irrigation (or micro-irrigation) or carry out manual watering to prevent wetting leaves, stems, and fruits
- Keep records of diseased species and times of occurrence. Symptoms of anthracnose often are seasonal, appearing with the natural cycle of senescence and disappearing with the flush of new growth.

Chemical

- Several fungicides are available that provide varying levels of control of anthracnose on ornamental plants. Fungicides can only protect healthy tissue and do not cure existing infections. During periods of wet, humid weather typical of the rainy season that favor spore dispersal and active infections, consider applying fungicides according to the shorter interval listed on the product label (usually 7 to 10 days). Spread fungicide applications over longer intervals when drier conditions are unfavorable for fungal infection. Plants showing resistance to or tolerance of anthracnose infection may not require fungicide applications or may only need a seasonal preventative treatment.
- Protective fungicides recommended for managing Colletotrichum spp. of ornamental plants contain chlorothalonil (Daconil Ultrex, etc.), Mancozeb (Manzate Max, etc.), or copper sprays containing copper diammonia diacetate or copper octanoate. The systemic fungicides thiophanate-methyl (Cleary's 3336, for professional use only), chlorothalonil + thiophanate methyl (Spectro 90WDG), pyraclostrobin + boscalid (Pageant, etc.), and tebuconazole (Tebuzol 3.6F, intended for professional applicators) offer the

greatest control if applied before disease development. For more information on other fungicide products and active ingredients, see EDIS publication PP154, "Homeowner's Guide to Fungicides for Lawn and Landscape Disease Management." Read and follow all label directions.

 Systemic fungicides are not recommended or labeled for edible plants in the landscape since the residual toxic compounds, or their metabolites, may persist in the edible fruit and vegetative structures. Some product labels specify that treated fruit should not be consumed (ornamental only). Read and follow all label directions carefully.



Figure 1. Spores of *C. gloeosporioides* under a compound light microscope.

Credit: Philip F. Harmon, UF/IFAS



Figure 2. *Colletotrichum* spp. with setae. Credit: Philip F. Harmon, UF/IFAS



Figure 3. Orange anthracnose hyphae erupting from the acervuli of *Collectotrichum gloeosporioides*.

Credit: Philip F. Harmon, UF/IFAS



Figure 4. *Agapanthus africanus* dieback of leaf tips. Credit: Stephen H. Brown, UF/IFAS



Figure 5. Anthracnose leaf blight of *Begoniα* spp. Credit: Stephen H. Brown, UF/IFAS



Figure 6. Lesions expanding along lateral leaf veins on *Begonia* spp.



Figure 7. Anthracnose blight on *Aspidistra elatior*. Credit: Stephen H. Brown, UF/IFAS



Figure 8. Necrotic spots usually begin at the leaf tips on crinum lily.

Credit: Stephen H. Brown, UF/IFAS



Figure 9. Yellowing of the leaves occurs with necrotic lesions on the desert rose.



Figure 10. Anthracnose of liriope. Credit: Stephen H. Brown, UF/IFAS



Figure 11. Anthracnose symptoms from the stem bottoms of bird of paradise.



Figure 14. Anthracnose symptoms on staghorn fern. Credit: Stephen H. Brown, UF/IFAS



Figure 12. Tree philodendron with blotches on leaf margins. Credit: Stephen H. Brown, UF/IFAS



Figure 15. Large blotch on an infected leaf of Awabuki viburnum.



Figure 13. *Thaumatophyllum* 'Xanadu' with marginal necrosis. Credit: Stephen H. Brown, UF/IFAS



Figure 16. Holes in a leaf of blue porterweed. Credit: Stephen H. Brown, UF/IFAS



Figure 17. Large blotches at the tips of cardboard palm leaflets.



Figure 18. Anthracnose blight on *Carissa macrocarpa*. Credit: Stephen H. Brown, UF/IFAS



Figure 19. Large anthracnose lesions are often accompanied by chlorotic leaves on Clusia sp. Credit: Stephen H. Brown, UF/IFAS



Figure 20. Necrotic leaf tips are typical symptoms on corn plants.

Credit: Stephen H. Brown, UF/IFAS



Figure 21. Codiaeum varigatum 'Petra' with marginal leaf chlorosis and necrosis.

Credit: Stephen H. Brown, UF/IFAS



Figure 22. Apical leaflet chlorosis and necrosis of dwarf schefflera.



Figure 23. Lesions concentrated in the tissue between the lateral leaf veins of firebush.



Figure 26. Anthracnose symptoms on 'Green Island' ficus. Credit: Stephen H. Brown, UF/IFAS



Figure 24. The death of entire stems occurring on foxtail ferns. Credit: Stephen H. Brown, UF/IFAS



Figure 27. Anthracnose leaf spots on $\it Hibiscus\ rose\alpha\mbox{-}sinensis.$ Credit: Stephen H. Brown, UF/IFAS



Figure 25. Leaf tip necrosis of *Duranta erecta* 'Gold Mound'. Credit: Stephen H. Brown, UF/IFAS



Figure 28. Indian hawthorn enlarged lesions. Credit: Stephen H. Brown, UF/IFAS



Figure 29. Infected leaves of *Ixora coccinea*. Credit: Stephen H. Brown, UF/IFAS



Figure 30. *Jatropha* sp. chlorosis and leaf spots. Credit: Ryan Czaplewski, UF/IFAS



Figure 31. Stages of leaf tip necrosis of jatropha. Credit: Stephen H. Brown, UF/IFAS



Figure 32. Chlorotic leaves with leaf spots on mandevilla. Credit: Stephen H. Brown, UF/IFAS



Figure 33. *Nerium oleander* stem dieback. Credit: Stephen H. Brown, UF/IFAS



Figure 34. *Nerium oleander* stem lesion. Credit: Stephen H. Brown, UF/IFAS



Figure 35. Necrosis at the leaf tips expands over the entire leaf of the ti plant.



Figure 36. Anthracnose infects leaflets along the length of this foxtail palm frond.

Credit: Stephen H. Brown, UF/IFAS



Figure 37. Leaf spots and the cupping of leaflets on foxtail palm.



Figure 38. Symptoms of anthracnose on *Bixa orellana*. Credit: Stephen H. Brown, UF/IFAS



Figure 39. Avocado fruit rot. Credit: Romina Gazis-Seregina, UF/IFAS



Figure 42. Mango fruit infected with anthracnose. Credit: Ian Maguire (emeritus), UF/IFAS



Figure 40. Leaf tip necrosis and blossom decay of lychee. Credit: Stephen H. Brown, UF/IFAS



Figure 43. Magnolia leaf blotches. Credit: Stephen H. Brown, UF/IFAS



Figure 41. Black spots with yellow halos on mango leaves. Credit: Stephen H. Brown, UF/IFAS



Figure 44. Pink trumpet tree with seasonal tip necrosis. Credit: Stephen H. Brown, UF/IFAS



Figure 45. Seasonal leaf tip necrosis of queen crape myrtle. Credit: Stephen H. Brown, UF/IFAS



Figure 46. Chlorotic and shedding pinnae of royal poinciana. Credit: Stephen H. Brown, UF/IFAS



Figure 47. Chlorotic blotches and spots along the leaf veins of strangler fig.



Figure 48. Defoliation of the strangler fig. Credit: Stephen H. Brown, UF/IFAS



Figure 49. Leaf tip necrosis of white bird of paradise. Credit: Stephen H. Brown, UF/IFAS



Figure 50. Irregular black spots with yellow halos on white mangrove.

References

- Crane, J. H., C. F. Balerdi, and I. Maguire. 2006. "Mango Growing in the Florida Home Landscape: HS2/MG216, rev. 10/2005." *EDIS* 2006 (18). https://doi.org/10.32473/edis-mg216-2006
- DeSilva, D. D., P. W. Crous, P. K. Ades, K. D. Hyde, P. W. J. Taylor. 2017. "Lifestyles of *Colletotrichum* Species and Implications for Plant Biosecurity." *Fungal Biology Reviews* 31 (3): 155–168. https://doi.org/10.1016/j.fbr.2017.05.001
- Douglas, S. M. 2011. Anthracnose Diseases of Trees. The Connecticut Agricultural Experiment Station. https://portal.ct.gov/-/media/caes/documents/publications/fact_sheets/plant_pathology_and_ecology/anthracnosediseasesoftrees060311rpdf.pdf
- Downer, A. J., S. S. Swain, and A. C. Crump. 2020. "Home and Landscape: Anthracnose." UC IPM. UC ANR Publication 7420. https://ipm.ucanr.edu/home-and-landscape/anthracnose/pest-notes/#gsc.tab=0
- Growables, Inc. 2013. "Avocado Diseases." Growables: Grow Florida Edibles. Last revised May 2, 2020. https://www.growables.org/information/Tropica lFruit/AvocadoDiseasesInsects.htm
- Nelson, S. 2008. "Anthracnose of Avocado." Publication No. PD-58. University of Hawaii at Manoa, Cooperative Extension Service. https://www.ctahr.hawaii.edu/oc/freepubs/pdf/pd-58.pdf
- Nelson, S. C. 2008. "Mango Anthracnose (Colletotrichum gloeosporioides)." Publication No. PD-48.
 University of Hawaii at Manoa, Cooperative Extension Service.
 https://www.ctahr.hawaii.edu/oc/freepubs/pdf/pd-48.pdf
- Pernezny, K., and R. B. Marlatt. 2007. "Diseases of Avocado in Florida." PP21. University of Florida Institute of Food and Agricultural Sciences. https://ufdcimages.uflib.ufl.edu/IR/00/00/28/56/00001/VH04700.pdf
- Popenoe, J., C. R. Warick, and D. J. Norman. 2021. "Key Plant, Key Pests: Lilyturf (*Liriope muscari*): EP600/ENH1336, 3/2021." *EDIS* 2021 (2). https://edis.ifas.ufl.edu/publication/EP600
- Weir, B. S., P. R. Johnston, U. Damm. 2012. "The Colletrotrichum gloeosporioides Species Complex." Studies in Mycology 73 (1): 115–180. https://doi.org/10.3114/sim0011

Table

Table 1. A partial list of landscape plants affected by *Colletotrichum* spp.

Type of plants	Common names	Botanical names
Herbaceous perennials	Agapanthus	Agapanthus africanus (Figure 4)
	Begonia	Begonia spp. (Figure 5, Figure 6)
	Cast iron plant	Aspidistra elatior (Figure 7)
	Crinum lily	Crinum asiaticum (Figure 8)
	Desert rose	Adenium obesum (Figure 9)
	Liriope	Liriope muscari (Figure 10)
	Orange bird of paradise	Strelitizia reginae (Figure 11)
	Tree philodendron	Thaumatophyllum bipinnatifidum (Figure 12)
	Xanadu	Thaumatophyllum xanadu (Figure 13)
Fern	Staghorn fern	Platycerium bifurcatum (Figure 14)
Shrubs	Awabuki viburnum	Viburnum odoratissimum 'Awabuki' (Figure 15)
	Blue porterweed	Stachytarpheta jamaicensis (Figure 16)
	Cardboard palm	Zamia furfuracea (Figure 17)
	Carissa	Carissa macrocarpa (Figure 18)
	Clusia	Clusia guittifera (Figure 19)
	Corn plant/Dracaena	Dracaena fragrans (Figure 20)
	Croton	Codiaeum variegatum (Figure 21)
	Dwarf schefflera	Schefflera arboricola (Figure 22)
	Firebush	Hamelia patens (Figure 23)
	Foxtail fern	Asparagus aethiopicus 'Myers' (Figure 24)
	Gold mound	Duranta erecta 'Gold Mound' (Figure 25)
	Green island ficus	Ficus microcarpa 'Green Island' (Figure 26)
	Hibiscus	Hibiscus rosa-sinensis (Figure 27)
	Indian hawthorn	Raphiolepis indica (Figure 28)
	Ixora	Ixora coccinea (Figure 29)
	Jatropha	Jatropha integerrima (Figure 30, Figure 31)
	Mandevilla	Mandevilla splendens (Figure 32)
	Oleander	Nerium oleander (Figure 33, Figure 34)
	Ti plant	Cordyline terminalis (Figure 35)
Palm	Foxtail palm	Wodyetia bifurcata (Figure 36, Figure 37)

Type of plants	Common names	Botanical names
Trees	Annatto	Bixa orellana (Figure 38)
	Avocado	Persea americana (Figure 39)
	Lychee	Litchi chinensis (Figure 40)
	Mango	Mangifera indica (Caution) (Figure 41, Figure 42)
	Magnolia	Magnolia spp. (Figure 43)
	Pink trumpet tree	Tabebuia heterophylla (Figure 44)
	Queen crape myrtle	Lagerstroemia speciosa (Figure 45)
	Royal poinciana	Delonix regia (Figure 46)
	Strangler fig	Ficus aurea (Figure 47, Figure 48)
	White bird of paradise	Strelitzia nicolai (Figure 49)
	White mangrove	Languncularia racemosa (Figure 50)

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² Stephen H. Brown, Extension agent IV, M.S., horticulture production, UF/IFAS Extension Lee County, Fort Myers, FL; Philip F. Harmon, professor and Extension plant pathologist, UF/IFAS Department of Plant Pathology, Gainesville, FL; Sheryl van der Heiden, Extension agent II, residential horticulture, UF/IFAS Extension Lee County, Fort Myers, FL; Ryan Czaplewski, interim county Extension director and Extension agent III, UF/IFAS Extension Collier County, Naples, FL; Ralph E. Mitchell, county Extension director and Extension agent IV, M.A., horticulture, UF/IFAS Extension Charlotte County, Port Charlotte, FL; David Outerbridge, county Extension director and program county Extension agent II, M.S., agriculture, UF/IFAS Extension Lee County, Fort Myers, FL; UF/IFAS Extension, Gainesville, FL 32611.