



The Hibiscus Bud Weevil (*Anthonomus testaceosquamosus* Linell, Coleoptera: Curculionidae)¹

Alexandra M. Revynthi, Yisell Velazquez-Hernandez, Juleysy Rodriguez, Paul E. Kendra, Daniel Carrillo, and Catharine M. Mannion²

Introduction

The hibiscus bud weevil (Anthonomus testaceosquamosus Linell, Coleoptera: Curculionidae) is a key insect pest of China rose hibiscus (Hibiscus rosa-sinensis L., Malvales: Malvaceae). This weevil originates from northeastern Mexico and southern Texas and was first found in Florida (in Miami-Dade County) in May 2017 (Skelley and Osborne 2018). It has since created severe hibiscus production and retail distribution challenges. According to ornamental nursery growers, increased weevil population densities in Homestead nurseries in 2019 and 2020 negatively impacted the hibiscus industry in south Florida during the spring shipping period, resulting in large economic losses. Florida leads hibiscus production nationally, and the majority of nursery production occurs in south Florida. Approximately 20%–25% of plants sold from Miami-Dade County are hibiscus, where the market value of ornamental plants was \$945,688 (farmgate price) in 2022 (United States Department of Agriculture 2022). To stop the spread and economic impact of the hibiscus bud weevil, the Florida Department of Agriculture and Consumer Services, Division of Plant Industry (FDACS-DPI) regulates this pest. Since the hibiscus bud weevil is a regulated pest, any nursery found with this weevil must sign and follow a compliance agreement with FDACS-DPI to reduce the chance of spreading the weevil. The purpose of this article is to provide nursery owners, homeowners, and other interested people with information for managing this serious pest.

Identification

The hibiscus bud weevil is a beetle (Order Coleoptera) in the weevil family Curculionidae. It belongs to the *Anthonomus squamosus* species-group of the Anthonomini tribe. This species-group is characterized by weevils predominantly covered with scales and includes several native and exotic weevil species (Clark et al. 2019) (Figure 1). The adult body length is 2.5 mm–2.7 mm (\sim 3/32 in) and the snout or beak is approximately 1 mm (1/32 in) long.



Figure 1. Adult *Anthonomus testaceosquamosus* Linell, a) lateral and b) dorsal habitus.

Credit: Daniel Carrillo, UF/IFAS TREC

Females can be distinguished from males using two characters, one in the protibia (the fourth segment of the first pair of legs) and one in the abdomen. In the protibia, females have an apical uncus and subapical, innermarginal prominence (mucron) (a spur-like structure on the inner side of the tibia) (Figure 2a), which is absent in males (Figure 2b). Additionally, the posterior part of the fifth ventrite (the margin of the fifth abdominal segment) in females is straight (Figure 3a, right) and in males is curved (Figure 3b, right). Moreover, females (Figure 3a, left) have a small pygidium (the last abdominal segment, exposed when the elytra are at rest) in comparison to males (Figure 3b, left). The validity of these characters was confirmed by dissecting weevil genitalia (Figure 4).



Figure 2. a) Female and b) male protibias of *Anthonomus* testaceosquamosus Linell. Subapical inner-marginal prominence (circled) (mucron) is present in females while absent in males.

Credit: Daniel Carrillo, UF/IFAS TREC

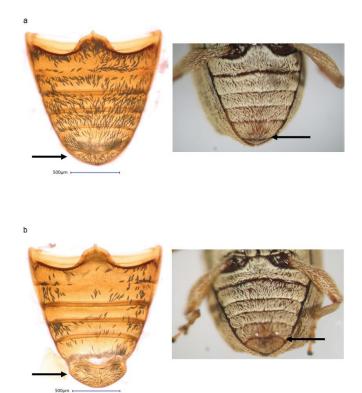


Figure 3. a) Female and b) male abdomens of *Anthonomus* testaceosquamosus Linell. The posterior end of the fifth ventrite in females is straight (a, right arrow) and in males is curved (b, right arrow). Females (a, left arrow) have a small pygidium (the last body part that is exposed when the elytra is at rest) in comparison to males (b, left arrow).

Credit: Daniel Carrillo, UF/IFAS TREC

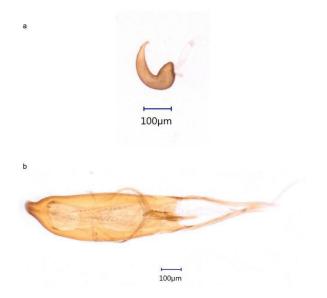


Figure 4. a) Female and b) male genitalia of *Anthonomus* testaceosquamosus Linell.

Credit: Daniel Carrillo, UF/IFAS TREC

Eggs (Figure 5), which are deposited inside a hibiscus flower bud, are white when newly deposited and turn yellow as they mature. The larvae of the hibiscus bud weevil are transparent to yellow, have a well-developed head capsule, and lack thoracic legs (Figure 6). The size of an individual larva is somewhat limited by the size of the

flower bud it occupies. In general, larger flower buds harbor larger larvae.



Figure 5. Females of *Anthonomus testaceosquamosus* Linell oviposit multiple eggs on hibiscus anthers within the flower bud.

Credit: Juleysy Rodriguez and Yisell Velazquez-Hernandez, UF/IFAS TREC





Figure 6. a) Early and b) late instar larvae of *Anthonomus testaceosquamosus* Linell feeding on pollen within the hibiscus flower bud.

Credit: Juleysy Rodriguez and Yisell Velazquez-Hernandez, UF/IFAS TREC

In Florida, another species from the *Anthonomus squamosus* species-group, *Anthonomus rubricosus*Boheman, has been reported infesting cotton and hibiscus plants (Clark et al. 2019; Loiácono et al. 2003). However, there are no reports showing its establishment in Florida. This weevil is similar in size to the hibiscus bud weevil but is brown. The *Anthonomus* genus includes several pest species of great agricultural importance, such as the cotton boll weevil, *Anthonomus grandis* Boheman. The most economically important *Anthonomus* pests reported in Florida are the pepper weevil, *Anthonomus eugenii* Cano, and the acerola weevil, *Anthonomus macromalus* Gyllenhal. The pepper weevil attacks plants of the family Solanaceae, particularly peppers (*Capsicum* spp.) (Capinera 2002), while the acerola weevil attacks Barbados cherry

(*Malpighia glabra*, Family: Malpighiaceae) (Hunsberger and Pena 1998).

Host Range and Damage

Weevils belonging to the *Anthonomus squamosus* speciesgroup are associated with plant species in the families, Asteraceae or Malvaceae. The hibiscus bud weevil, *Anthonomus testaceosquamosus*, has been associated with 15 plant species, all from the family Malvaceae (Table 1).

Adult weevils feed primarily on hibiscus buds, stems, and, to a lesser extent, on the leaves. The females oviposit in flower buds, and larvae develop inside the bud, causing bud drop prior to flowering. Infestation symptoms include holes in stems and unopened buds (Figure 7) and severe bud drop when weevil densities are high. Hibiscus bud weevil infestation has been associated with a 16-fold increase in flower bud abortion compared to non-infested plants (Vargas et al. 2024). Feeding damage on the leaves is not apparent. In south Florida nurseries, pink and yellow hibiscus varieties appear to be the most susceptible to hibiscus bud weevil (Table 2). More specifically, the pink variety 'Painted Lady' and the yellow variety 'Sunny Yellow' are reported to be the most susceptible varieties. The red variety 'President Red' is reported to be the most tolerant to hibiscus bud weevil.





Figure 7. Feeding damage caused by *Anthonomus* testaceosquamosus Linell on hibiscus; a) bud with adult weevil and b) petiole with signs of infestation.

Credit: Juleysy Rodriguez and Yisell Velazquez-Hernandez,

UF/IFAS TREC

Another pest causing hibiscus flower bud drop is the hibiscus bud midge (*Contarinia maculipennis*, Diptera: Cecidomyiidae), which can be mistaken for hibiscus bud weevil damage (Mannion et al. 2006). Both pests can infest the same hibiscus plant; however, they will rarely infest the same flower bud. Buds infested with the hibiscus bud midge will have multiple white to yellow maggots (larvae) inside them that jump when disturbed. Midge maggots lack a head and legs and need to exit the bud to pupate in the soil while the weevil larvae have true heads and pupate within the buds (Figure 8; Figure 9).



Figure 8. Larva of a) hibiscus bud weevil (*Anthonomus testaceosquamosus* Linell) and b) hibiscus bud midge (*Contarinia maculipennis* Felt).

Credit: Juleysy Rodriguez and Yisell Velazquez-Hernandez, UF/IFAS TREC



Figure 9. A hibiscus bud midge (*Contarinia maculipennis* Felt) larva exits the flower bud. This photo shows the damage caused on the flower bud by the feeding maggots.

Credit: Juleysy Rodriguez and Yisell Velazquez-Hernandez,

UF/IFAS TREC

Biology

Adult female weevils deposit 3–5 eggs into a single hibiscus flower bud close to the anthers (Figure 5). Emerged larvae feed on pollen and remain in the flower bud until they reach adulthood. Due to the high incidence of larval cannibalism, not all eggs will successfully develop into an adult weevil; however, multiple weevils may exit one flower bud. At 27°C (80.6°F), eggs hatch within 2–3 days of oviposition. The larval stage has three instars and can last, on average, 10 days. The pupal stage (Figure 10) lasts 2.9–4.2 days. Development from egg to adult can range from 12.8–16.3 days, and survival has been observed up to 90%. Adult longevity ranges from 13–169 days, with the males living longer than the females. When fed solely on pollen, adults can survive up to 52 days. Adult weevils

can survive on an average for 28 days without food when they have access to water, and 16 days without water. The sex ratio is 1:1 females to males (Revynthi et al. 2022). Preliminary research has shown that the hibiscus bud weevil can also complete its life cycle on okra flower buds (*Abelmoschus esculentus*).



Figure 10. Pupa of *Anthonomus testaceosquamosus* Linell. Credit: Juleysy Rodriguez and Yisell Velazquez-Hernandez, UF/IFAS TREC

Very low (<10°C or 50°F) or high (>35°C or 95°F) temperatures appear to be detrimental to the development of this weevil. In laboratory experiments at the University of Florida, at 10°C (50°F) no eggs hatched, while at 15°C (59°F) eggs hatched 12 days after oviposition, but the larvae did not feed and eventually died. Similarly, at 34°C (93.2°F), eggs hatched within 5.5 days, but none of the larvae managed to pupate. In south Florida, the peak activity of this weevil has been observed from March through June (23°C –28°C or 73.4°F –82.4°F) with lower populations from September until February (22°C –28°C or 71.6°F –82.4°F). Based on this information and high dependence on the hibiscus plant, the weevil is expected to be limited to tropical and subtropical areas with large hibiscus production.

Development of Pest Management and Monitoring Techniques

Integrated pest management programs targeting the hibiscus bud weevil should involve a combination of cultural practices, sanitation, active monitoring, chemical intervention, and biological control. Implementing a combination of these practices will reduce reliance on insecticide inputs for plant protection.

Cultural Practices

Crop rotation with non-host species has been recommended for interrupting population cycles by removing host plant material and insect reservoirs within production settings (Bográn et al. 2003). Sanitation includes systematic collection and destruction of all dropped buds from the ground. Although sanitation is labor-intensive, it has been proposed as the most efficient management practice against this weevil since dropped buds contain weevils that will continue to infest new plants (Bográn et al. 2003). A weekly collection of dropped buds resulted in a 22% reduction in the proportion of dropped buds in comparison to plants where no sanitation was implemented (Vargas et al. 2024). The most time-efficient method to implement sanitation is to use a vacuum (Vargas et al. 2024).

Chemical Control

Contact-toxic insecticides target the adult weevils that live freely on the hibiscus plant canopy, while systemic insecticides target the developing hibiscus bud weevil larvae inside the flower buds. Laboratory and greenhouse experiments evaluating the efficacy of contact-toxic insecticides showed that diflubenzuron (Dimilin® 25W, 16 oz/acre, IRAC group15, ohp); pyrethrins (PyGanic Crop Protection® EC, 15.61 fl oz/acre, IRAC group 3A, Valent); spinetoram plus sulfoxaflor (Xxpire®, 2.75 oz/acre, IRAC group 4C+5, Corteva); and spirotetramat (Kontos®, 3.4 fl oz/acre, IRAC group 23, Envu) provided the highest level of control. Moreover, mineral oils (Ultrafine®, 2%, unknown IRAC group, HollyFrontier and Suffoil-X®, 2%, unknown IRAC group, BioWorks) can also be effective contact insecticides targeting adult weevils (Greene et al. 2023). Applications of the non-neonicotinoid systemic insecticides chlorantraniliprole (Acelepryin®, 16 fl oz/acre, IRAC group 28, Syngenta); flupyradifurone (Altus®, 28 fl oz/acre, IRAC group 4D, Envu); and cyantraniliprole (Mainspring®, 12 fl oz/acre, IRAC group 28, Syngenta) one month before a hibiscus infestation limited weevil feeding and oviposition activity and reduced larval survival (Vargas et al. 2023).

Biological Control

Biological control using entomopathogenic nematodes can offer a sustainable and environmentally friendly solution for managing the hibiscus bud weevil. Foliar spray applications of *Steinernema carpocapsae* resulted in significant weevil larval mortality within hibiscus flower buds. In addition, this foliar spray nematode application can also control weevil larvae in hibiscus flower buds that have fallen to the soil surface beneath potted plantings (Vargas et al. 2024). Entomopathogenic fungi and bacteria can be used to target adult hibiscus bud weevils. Research shows that the fungus *Beauveria bassiana* and the bacterium *Chromobacterium subtsugae* have the potential to reduce weevil reproduction and feeding. Recently, a polyphagous ectoparasitoid wasp, *Catolaccus hunteri* Crawford (Hymenoptera: Pteromalidae), became available

in the American commercial biological control market. This tiny wasp parasitizes closely related pest weevils, the cotton boll weevil and the pepper weevil (Vazquez 2004; Gomez-Dominguez et al. 2012). Preliminary studies have shown that *Catolaccus hunteri* can also parasitize hibiscus bud weevil larvae, warranting further research.

Pest Monitoring Tools

Pheromone traps broadly used for other Anthonomus species are being explored as a pest-monitoring tool. Several species within the Anthonomus genus are attracted to a group of commercial lures that contain male aggregation pheromones and host plant volatiles that are attractive to adult weevils (Tumlinson et al. 1969; Eller et al. 1994; Innocenzi et al. 2001). There are four components of the synthetic male aggregation pheromone, also known as Grandlures I-IV. In Texas, cotton boll weevil pheromone traps were evaluated and determined to not successfully attract hibiscus bud weevil adults, although this requires further investigation (Bográn et al. 2003). Yellow sticky traps are the most attractive traps for several Anthonomus species, including the hibiscus bud weevil (Cross et al. 2006; Szendrei et al. 2011; Silva et al. 2018; Ataide et al. 2024). Trials conducted to test hibiscus bud weevil attraction to lures for boll, pepper, and cranberry weevils (Anthonomus musculus) showed that yellow sticky cards with the cranberry weevil lure placed at canopy height caught the largest number of adult hibiscus bud weevils (Ataide et al. 2024).

The best way to manage the hibiscus bud weevil in Florida and decrease the economic damage it causes is to use an IPM program implementing all of the strategies described above. Monitor hibiscus crops each week to detect weevils early. Sanitize to break the weevil population cycle by collecting and destroying dropped hibiscus buds at least twice per month throughout the hibiscus growing season (August-March). When weevil populations are low (August-February), employ biological control agents such as entomopathogenic nematodes, fungi, or bacteria to help keep populations under control, reducing the need for contact insecticide applications. As the plants grow and produce more flower buds (around February), systemic insecticide applications can offer protection from increasing weevil populations. As the shipping season approaches (March-May), to comply with existing regulations, contact insecticides may be applied to ensure effective control and production of clean plant material.

References

Ataide, L. M. S., A. D. Greene, K. R. Cloonan, et al. 2024.

"Exploring Market-Available Pheromone Lures and Traps for Monitoring

Anthonomustestaceosquamosus (Coleoptera: Curculionidae)." Journal of Economic Entomology 117 (10): 1396–1405.

https://doi.org/10.1093/jee/toae105

- Bográn, C. E., K. M. Heinz, and S. Ludwig. 2003. "The Bud Weevil *Anthonomus testaceosquamosus*, a Pest of Tropical Hibiscus." In: SNA Research Conference Entomology. pp 147–149.
- Capinera, J. L. 2002. "Pepper Weevil Anthonomus eugeniiCano (Insecta: Coleoptera: Curculionidae)."

 EENY-278. Gainesville: University of Florida
 Institute of Food and Agricultural Sciences.

 https://edis.ifas.ufl.edu/publication/IN555
- Clark, W. E., H. R. Burke, R. W. Jones, and R. S. Anderson. 2019. "The North American Species of the *Anthonomus squamosus* Species-Group (Coleoptera: Curculionidae: Curculioninae: Anthonomini)." *Coleopterists Bulletin* 73 (4): 773. https://doi.org/10.1649/0010-065X-73.4.773
- Cortez-Mondaca, E., N. M. Bárcenas-Ortega, J. L Martínez-Carrillo, et al. 2004. "Parasitismo de *Catolaccus grandis y Catolaccus hunteri* (Hymenoptera: Pteromalidae) Sobre el Picudo del Algodonero (*Anthonomus grandis* Boheman)." *Agrociencia* 38 (5): 497–501.
- Cross, J. V., H. Hesketh, C. N. Jay, et al. 2006. "Exploiting the Aggregation Pheromone of Strawberry Blossom Weevil Anthonomus rubi Herbst (Coleoptera: Curculionidae): Part 1. Development of Lure and Trap." Crop Protection 25:144–154. https://doi.org/10.1016/j.cropro.2005.04.002
- Eller, F. J., R. J. Bartelt, B. S. Shasha, et al. 1994.

 "Aggregation Pheromone for the Pepper Weevil,

 Anthonomus eugenii Cano (Coleoptera:

 Curculionidae): Identification and Field Activity."

 Journal of Chemical Ecology 20:1537–1555.

 https://doi.org/10.1007/BF02059879
- Gomez-Dominguez, N. S., R. J. Lomeli-Flores, E. Rodriguez-Leyva, J. M. Valdez-Carrasco, and A. Torres-Ruiz. 2012. "Ovipositor of *Catolaccus hunteri* Burks (Hymenoptera: Pteromalidae) and Implications for its Potential as a Biological Control Agent of Pepper Weevil." Southwestern Entomologist 37 (2): 239–242. https://doi.org/10.3958/059.037.0218
- Greene, A. D., X. Yang, Y. Velazquez-Hernandez, et al. 2023. "Lethal and Sublethal Effects of Contact Insecticides and Horticultural Oils on the Hibiscus Bud Weevil, *Anthonomustestaceosquamosus* Linell (Coleoptera: Curculionidae)." *Insects* 14 (6): 544. https://doi.org/10.3390/insects14060544

- Hunsberger, A., J. E. Pena, R. M. Giblin-Davis, G. Gries, and R. Gries. 1998. "Biodynamics of *Anthonomus macromalus* (Coleoptera: Curculionidae), a Weevil Pest of Barbados Cherry in Florida." Florida State Horticultural Society 111:334–338.
- Innocenzi, P. J., D. R. Hall, and J. V. Cross. 2001.

 "Components of Male Aggregation Pheromone of Strawberry Blossom Weevil, *Anthonomus rubi*Herbst. (Coleoptera: Curculionidae)." *Journal of Chemical Ecology* 27:1203–1218.

 https://doi.org/10.1023/A:1010320130073
- Loiácono, M. S., A. E. Marvaldi, and A. A. Lanteri. 2003. "Description of Larva and New Host Plants for *Anthonomus rubricosus* Boheman (Coleoptera: Curculionidae) in Argentina." *Entomological News* 114:69–74.
- Mannion, C., A. Hunsberger, K. Gabel, E. Buss, and L. Buss. 2006. Hibiscus bud midge (*Contarinia maculipennis*). UF/IFAS Extension. http://trec.ifas.ufl.edu/mannion/pdfs/HibiscusMidge.pdf (no longer available online).
- Revynthi, A. M., Y. V. Hernandez, M. A. Canon, et al. 2022. "Biology of *Anthonomustes taceosquamosus* Linell, 1897 (Coleoptera: Curculionidae): A New Pest of Tropical Hibiscus." *Insects* 13 (1): 1–14. https://doi.org/10.3390/insects13010013
- Silva, D., J. Salamanca, V. Kyryczenko-Roth, et al. 2018.

 "Comparison of Trap Types, Placement, and Colors for Monitoring *Anthonomus musculus* (Coleoptera: Curculionidae) Adults in Highbush Blueberries."

 Journal of Insect Science 18 (2): 19; 1–9.

 https://doi.org/10.1093/jisesa/iey005
- Skelley, P. E., and L. S. Osborne. 2018. *Pest Alert*Anthonomus testaceosquamosus Linell, the Hibiscus

 Bud Weevil, New in Florida. FDACS-P-01553.

 Florida Department of Agriculture and Consumer

 Services Division of Plant Industry.
- Szendrei, Z., A. Averill, H. Alborn, and C. Rodriguez-Saona. 2011. "Identification and Field Evaluation of Attractants for the Cranberry Weevil, *Anthonomus musculus* Say." *Journal of Chemical Ecology* 37:387–397. https://doi.org/10.1007/s10886-011-9938-z
- Tumlinson, J. H., D. D. Hardee, R. C. Gueldner, et al. 1969. "Sex Pheromones Produced by Male Boll Weevil: Isolation, Identification, and Synthesis." *Science* 166 (3908): 1010–1012. https://doi.org/10.1126/science.166.3908.1010

- United States Department of Agriculture. 2022. "Table 2.

 Market Value of Agricultural Products Sold
 Including Food Marketing Practices and ValueAdded Products: 2022 and 2017" In the 2022
 Census of Agriculture Volume 1, Chapter 2: State
 Level Data.
 https://www.nass.usda.gov/Publications/AgCens
 us/2022/Full_Report/Volume_1,_Chapter_2_US_St
 ate_Level/
- Vargas, G., A. D. Greene, Y. Velazquez-Hernandez, et al. 2023. "A Prophylactic Application of Systemic Insecticides Contributes to the Management of the Hibiscus Bud Weevil Anthonomus testaceosquamosus Linell (Coleoptera: Curculionidae)." Agriculture 13 (10): 1879. https://doi.org/10.3390/agriculture13101879
- Vargas, G., A. Daniel Greene, Y. Velazquez-Hernandez, et al. 2024. "Implementing Sanitation Practices Against the Hibiscus Bud Weevil Anthonomus testaceosquamosus Linell (Coleoptera: Curculionidae)." Florida Entomologist 107 (1) 2024, pp. 20240023. https://doi.org/10.1515/flaent-2024-0023
- Vargas, G., Y. Velazquez-Hernandez, A. Daniel Greene, et al. 2024. "Entomopathogenic Nematodes to Control the Hibiscus Bud Weevil *Anthonomus testaceosquamosus* (Coleoptera: Curculionidae), Above Ground and on Soil Surface." *BioControl* 69:1–11. https://doi.org/10.1007/s10526-024-10242-9

Table 1. Plant species on which the hibiscus bud weevil *Anthonomus testaceosquamosus* Linell has been found (Clark et al. 2019).

Plant species scientific name	Plant common name
Abutilon abutiloides	shrubby Indian mallow
Abutilon simulans	corona de reina
Hibiscus acicularis	rose mallow
Hibiscus cardiophyllus	heartleaf hibiscus
Hibiscus martianus	tulipan del monte
Hibiscus rosa-sinesis	China rose hibiscus
Malvastrum americanum	Indian valley false mallow
Malvastrum corchorifolium	false mallow
Malvastrum spicatum	false mallow
Malvaviscus drummondii	Turk's cap mallow
Pseudabutilon hypoleucum	white-leaf Indian mallow
Pseudabutilon lozanii	Lozano's false Indian mallow
Sida sp.	fanpetals
Tillandsia ionantha	blushing bride
Wissadula holosericea	Chisos Mountain false Indian mallow

Table 2. Hibiscus varieties cultivated in Florida that have been found infested with the hibiscus bud weevil (*Anthonomus testaceosquamosus Linell*).

Hibiscus Variety	Flower Color
Double Orange	orange
Double Peach	
Orange Sunset	
Painted Lady	pink
Passion Wind	
Pink	
Salmon	
Seminole Pink	
Double red	red
Fiesta	
Hot Shot	
President Red	
Starry Wind	
Chatty	yellow
Fort Mayer's Yellow	
Sunny Yellow	
Tequila	
Yellow	

The Institute of Food and Agricultural Sciences (IFAS) is an Equal Opportunity Institution authorized to provide research, educational information and other services only to individuals and institutions that function with non-discrimination with respect to race, creed, color, religion, age, disability, sex, sexual orientation, marital status, national origin, political opinions or affiliations. For more information on obtaining other UF/IFAS Extension publications, contact your county's UF/IFAS Extension office. U.S. Department of Agriculture, UF/IFAS Extension Service, University of Florida, IFAS, Florida A & M University Cooperative Extension Program, and Boards of County Commissioners Cooperating. Andra Johnson, dean for UF/IFAS Extension.

¹This document is ENY-2069, one of a series of the Department of Entomology and Nematology, UF/IFAS Extension. Original publication date August 2021. Revised August 2025. Visit the EDIS website at https://edis.ifas.ufl.edu for the currently supported version of this publication. © 2025 UF/IFAS. This publication is licensed under CC BY-NC-ND 4.0

² Alexandra M. Revynthi, assistant professor, agricultural acarology, Department of Entomology and Nematology, UF/IFAS Tropical Research and Education Center; Yisell Velazquez-Hernandez, biological scientist II, ornamental plant entomology, UF/IFAS Tropical Research and Education Center; Juleysy Rodriguez, Department of Entomology and Nematology, UF/IFAS Tropical Research and Education Center; Paul E. Kendra, research entomologist, Subtropical Horticulture Research, USDA Agricultural Research Service; Daniel Carrillo, associate professor, Department of Entomology and Nematology, UF/IFAS Tropical Research and Education Center; Catharine M. Mannion, professor emeritus, ornamental entomology, Department of Entomology and Nematology; UF/IFAS Tropical Research and Education Center; UF/IFAS Extension, Gainesville, FL 32611.