

Saddleback Caterpillar *Acharia stimulea* (Clemens) (Insecta: Lepidoptera: Limacodidae)¹

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Introduction

Acharia stimulea (Clemens) is a limacodid moth, or slug moth, best known for its larval growth phase. Distinct bright color patterns and the presence of venomous, urticating spines lead to its recognition as the saddleback caterpillar. It is native to a large range in the eastern United States and able to feed on a wide array of host plant species. This species can survive well in northern temperate areas and warmer southern climates. The saddleback caterpillar is encountered most frequently as a medically significant pest, and has minor effects in landscaping and agriculture.

Synonymys

Empretia stimulea Clemens

Limacodes ephippiatus Harris

Sibine stimulea (Clemens)

Acharia stimulea (Clemens)

(Dyar and Morton 1896)



Figure 1. Mature larvae of the saddleback caterpillar, *Acharia stimulea* (Clemens).

Credits: Lyle J. Buss, University of Florida

Distribution

Acharia stimulea has a wide range in the eastern United States, occurring as far southward as Florida, northward to New York and Massachusetts, and westward to Texas, Indiana, and Kansas (Snow 1875; Darlington 1952; Nielsenbaum 1992; Landau and Prowell 1999; Heppner 2003; Covell 2005; Wagner 2005).

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Figure 2. State level distribution of *Acharia stimulea* (Clemens) in United States.

Description

Adults

Acharia stimulea adults are glossy dark brown in color with black shading. Dense scales are present on the body and wings, giving it a furry appearance. Wing span ranges from 26–43 mm (1.0–1.7 in), with females typically larger in size than males. A single white dot is present near the forewing base. Near the forewing apex, one to three additional white dots are present. The hind wings will be a paler brown than the forewings.



Figure 3. Adult stage of the saddleback caterpillar, *Acharia stimulea* (Clemens), showing the white dots on the wings.
Credits: Lyle J. Buss, University of Florida

Eggs

Clutches are laid on the upperside of host plant leaves in irregular clusters of thirty to fifty eggs. The eggs are sometimes laid to partially overlap each other. Individual eggs are yellow and transparent, with scale-like characteristics and thin edges. Length ranges from 1.5–2.0 mm (0.06–0.08

in) and width averages 1 mm (0.04 in). No additional markings are present.

Larvae

The caterpillars have a truncated, slug-like body form that is not typical of other Lepidoptera families. The prolegs are concealed under the ventral surface of the body. Color patterns are aposematic, or having bright warning colors that denote toxicity or distastefulness. The caterpillars are dark brown on the anterior and posterior ends with a contrasting bright green pattern blanketing the dorsal midsection.



Figure 4. Late instar larva of the saddleback caterpillar, *Acharia stimulea* (Clemens) larva, on blueberry.
Credits: Lyle J. Buss, University of Florida

This green section is bordered with white markings that outline a saddle-like shape. The skin has a granulated texture, and large fleshy tubercles extend from both anterior and posterior ends. Each tubercle is covered in long setae and a complex of varied urticating spines embedded among the setae.



Figure 5. Close up of urticating spines of the saddleback caterpillar, *Acharia stimulea* (Clemens) larva, on blueberry.
Credits: Lyle J. Buss, University of Florida

A variable marking occurs at the posterior end where a cream colored set of spots imitates a large facial area.



Figure 6. Close up the startle display marking of the saddleback caterpillar, *Acharia stimulea* (Clemens).
Credits: Lyle J. Buss, University of Florida

At hatching, larvae can be as small as 1.2 mm (0.05 in) and during last instar the larvae can be as large as 20 mm (0.8 in) with a 7 mm (0.28 in) width. Early instars lack defined aposematic coloration, but still have the stout body design and fleshy tubercles shown by later instars.



Figure 7. Early instar larvae of the saddleback caterpillar, *Acharia stimulea* (Clemens).
Credits: Lyle J. Buss, University of Florida

Middle instars begin to develop the well defined markings of the species and the enlarged spinal processes.



Figure 8. Middle instar larvae of the saddleback caterpillar, *Acharia stimulea* (Clemens).
Credits: Lyle J. Buss, University of Florida

Pupae

Larvae spin compact, silk cocoons roughly half the size of the last instar. The cocoon is then hardened to protect the pupa contained within.



Figure 9. Cocoon of the saddleback caterpillar, *Acharia stimulea* (Clemens).
Credits: Photograph by: Pennsylvania Department of Conservation and Natural Resources; Bugwood.org

A cocoon from which an adult has emerged will have a circular exit hole as if a hatch has opened. This cocoon design is typical of Limacodidae as a family, and not specific to *A. stimulea*.

Life Cycle and Biology

The transparent eggs reveal the small larvae of *A. stimulea*, which are observed curled inside with anterior and posterior ends touching (Dyar and Morton 1896; Epstein 1996;

Wagner 2005). After ten days, the minute larva will chew itself free from the egg, creating a small exit hole (Dyar and Morton 1896).

Larvae need approximately four to five months to feed and develop before pupating (Dyar and Morton 1896; Hossler 2010). The first instars lack the aposematic colors or startle display markings that complement their spine defenses. Hatchlings will feed gregariously on a single leaf until advancement to the next growth stage.

The second through penultimate instars acquire brown colorations on the anterior and posterior ends, and develop the bright green saddle markings on the midsection. Although still small at 5–8 mm (0.2–0.3 in) in length, they acquire urticating spines on the fleshy tubercles of the body as early as the second instar. The spinal processes continue to develop and fleshy horns become apparent on the anterior and posterior ends, which function as the primary location of defensive spines. The second to fourth instars feed indiscriminately on the upper or under sides of a host plant leaf, and feed on plant tissue between leaf veins while avoiding the leaf veins themselves; this creates visual dark patches (or ‘windows’) (Packard 1893; Dyar and Morton 1896).



Figure 10. Example of middle instar saddleback caterpillars, *Acharia stimulea* (Clemens), feeding in ‘windows’ on underside of leaf.
Credits: Lyle J. Buss, University of Florida

The last instars engorge themselves and reach lengths of approximately 20 mm (0.8 in) with a 7 mm (0.28 in) width. This fully fed stage has noticeably enlarged anterior and posterior ends, with well developed spinal processes and several fleshy horns. The last instars feed on the entire leaf, consuming veins, under side, and upper side regardless of feeding orientation on the leaf (Dyar and Morton 1896).

Saddleback caterpillars have modified prolegs where instead of crochets, which hook into materials and surfaces

to provide support, the limacodids have suckers (Epstein 1996). Larvae also secrete a semi-fluid silk from their ventral pores as they move. This helps increase body contact with their substrate and provides an adhesive bond, especially to smooth surfaces like those of palm plant leaves (Epstein 1996; Howard et al. 2001). This locomotive complex gives the family its designation as slug caterpillars because of their gliding movement pattern (Epstein 1996).



Figure 11. Last instar saddleback caterpillar, *Acharia stimulea* (Clemens), feeding on whole leaf.
Credits: Lyle J. Buss, University of Florida

When on the verge of pupation, limacodid caterpillars expel chalky white frass pellets (Epstein 1996). Fluid is also expelled from the body to reduce the body volume to approximately one half the size of the fully fed larva, at which point the color dulls and pigment is lost (McNaulty 1967). The expelled fluids help compress setae and spines to aid in compaction (Epstein 1996). The silk layer is modified with a circular weakened area at one end (Epstein 1996). This weakened area is not visible before eclosion, and forms a hatch that ejects when pressure is applied by the pupa (Dyar and Morton 1896; Common 1990; Schremmer 1990; Epstein 1996). Despite defensive measures in the cocoon, predation on the pupae accounts for a significant level of mortality in *A. stimulea* (Murphy and Lill 2010). Caterpillars in temperate areas will overwinter as pupae (USAF 1982; Epstein 1996; Murphy and Lill 2010).

Adults emerge for mating flights in June and July in far northeastern states (Dyar and Morton 1896; USAF 1982), which translates into expected emergences closer to February and March in warmer climate zones as in Texas and Florida. Males fly first in the twilight hours of warm nights, with females following during the night hours; successful couplings will stay in copulatory attachment for up to 24 hours (Dyar and Morton 1896). Adults have a single brood,

with oviposition occurring at night. *Acharya stimulea* is polyphagous, especially on deciduous trees (Scott 1964).

Hosts

What host plant this insect is found on is not a relevant means of identifying *A. stimulea* because of its polyphagous nature. The following list indicates known hosts:

Aceraceae: *Acer* spp., maple

Adoxaceae: *Viburnum* spp.

Anacardiaceae: *Schinus terebinthifolius*, Brazilian peppertree

Aquifoliaceae: *Ilex cornuta*, holly

Araceae: *Anthurium andraeanum*, flamingo flower; *Epipremnum* spp., pothos vine; *Philodendron selloum*

Arecaceae: *Areca* spp., single stemmed palms; *Archontophoenix alexandrae*, Alexander palm; *Adonidia merrilli*, Christmas palm; *Caryota mitis*, fishtail palm; *Chamaedorea* spp.; *Cocos nucifera*, coconut palm; *Dictyosperma album*, princess palm; *Dypsis lutescens*, butterfly palm; *Nannorrhops ritchiana*, mazari palm; *Phoenix canariensis*, Canary Island date palm; *Phoenix roebeleni*, pygmy date palm; *Syagrus romanzoffianum*, queen palm; *Washingtonia robusta*, Mexican fan palm

Asparagaceae: *Beaucarnea recurvata*, ponytail palm; *Cordyline terminalis*, cabbage tree; *Dracaena deremensis*, green dracaena; *D. fragrans* cornstalk dracaena; *D. marginata*, dragon tree

Asteraceae: *Aster* spp.; *Helianthus* spp., sunflower and artichoke

Bignoniaceae: *Tabebuia aurea* Caribbean trumpet tree; *T. umbellata* Ipê

Cannabaceae: *Celtis* spp., hackberry

Cannaceae: *Canna indica*, Indian shot

Cornaceae: *Cornus* spp., dogwood

Cycadaceae: *Cycas revoluta*, cycad, sago 'palm'

Ericaceae: *Vaccinium* spp., blueberry, cranberry, huckleberry, lingonberry

Euphorbiaceae: *Acalypha wilkesiana*; *Codiaeum* spp.

Fabaceae: *Wisteria* spp.

Fagaceae: *Castanea* spp., chestnut; *Quercus* spp., oak

Hydrangeaceae: *Hydrangea* spp.

Iridaceae: *Gladiolus* spp.

Juglandaceae: *Carya illinoensis*, pecan

Lauraceae: *Lindera benzoin*, spicebush

Leeaceae: *Leea coccinea*

Lythraceae: *Lagerstroemia indica*, crape myrtle

Malpighiaceae: *Malpighia glabra*, Barbados cherry

Malvaceae: *Hibiscus rosa-sinensis*, Chinese hibiscus; *H. syriacus*, rose of Sharon; *Malvaviscus* spp., wax mallow; *Tilia* spp., basswood

Meliaceae: *Swietenia mahogany*, West Indies mahogany

Moraceae: *Ficus elastica*, rubber fig

Myrtaceae: *Eucalyptus* spp.; *Feijoa sellowiana*, pineapple guava; *Melaleuca quinquenervia*, broad leaved paperbark; *Psidium cattleianum*, strawberry guava; *Syzygium* spp., brush cherry

Orchiaceae: *Cattleya* spp., orchid; *Chysis bractescens*; *Cyrtopodium punctatum*, cyrt; *Dendrobium nobile*, noble dendrobium; *Epidendrum tampense*; *Spathoglottis plicata*, large purple orchid

Poaceae: *Zea mays*, sweet corn

Podocarpaceae: *Podocarpus* spp.

Polygonaceae: *Antigonon leptopus*, coral vine; *Coccoloba uvifera*, seagrape

Proteaceae: *Macadamia tetraphylla*, macadamia nut

Rosaceae: *Eriobotrya japonica*, loquat; *Malus pumila*, common apple; *M. sylvestris*, crabapple; *Photinia* spp.; *Prunus serotina*, black cherry; *Pyrus* spp., pear; *Rosa* spp., rose

Rubiaceae: *Coffea arabica*, mountain coffee; *Gardenia* spp.; *Ixora coccinea*, jungle geranium

Rutaceae: *Citrus aurantium*, bitter orange; *C. limonia*, rangpur; *C. paradisi*, grapefruit; *C. sinensis*, sweet orange; *Fortunella* spp., kumquat

Sapindaceae: *Litchi chinensis*, lychee or litchi

Sapotaceae: *Pouteria campechiana*, canistel; *Sideroxylon lanuginosum*, false buckthorn

Strelitziaceae: *Strelitzia nicolai*, great white bird of paradise

Ulmaceae: *Ulmus* spp., elm

Vitaceae: *Vitis* spp. grapevine

(List compiled using Snow 1875; Darlington 1952; Cassani et al. 1989; Niesenbaum 1992; Howard et al. 2001; Costello et al. 2003; Hepner 2003)

Defenses

The contrasting dark brown ends and bright green saddle are a form of aposematism, which is common among organisms with strong defensive characteristics (Speed and Ruxton 2005). Not necessarily related to aposematism, there are also sets of cream colored spots on the posterior end of the caterpillar that seemingly imitates a face (Dyar and Morton 1986; Packard 1983). These spots are akin to those of other lepidopteran larvae that have more articulate eye spots or snake mimicry in that they confuse aggressors by seeming to have large eyes. These markings discourage predators from attacking by signaling danger (Speed and Ruxton 2005).

The family Limacodidae is also recognized for its members having fleshy horns on the body, each containing numerous hollow spines. In *A. stimulea*, these hollow spines are present on the edges of the body length, but are concentrated on tuberculated protrusions that adorn both ends of the larvae creating a total of four large horns. Spinose processes vary in shape, length, and articulation relative to where they are on the body, and whether or not multiple types are in proximity to each other (Dyar and Morton 1986; Gilmer 1925; Packard 1983). All of them are capable of breaking away from the body and remaining embedded in contacted surfaces. These spines contain hemolytic and vesicating venom, secreted from nearby glands, that causes direct tissue damage (Scott 1963; Edwards et al. 1986; Murphy et al. 2009).

Saddleback larvae use their great adhesive properties to stay firmly mounted to a surface. When coupled with the venomous spines on the body, this allows the caterpillar to

remain attached to any selected surface and offer its spines to aggressors. Sometimes, if disturbed, larvae will arch their back inward in an attempt to create contact with all anterior and posterior horns (Packard 1893; Murphy et al. 2009); however, they are unable to retract the head capsule into the body for protection (Packard 1893).

The aposematic signaling and venomous spines function together to preserve the caterpillar, as the metabolic cost of developing either characteristic is steep. By having both, the organism can enhance the effects of both characteristics and further decrease the probability of aggressive contact (Speed and Ruxton 2005). The large, potent spines of *A. stimulea* have been shown to effectively deter vertebrate and invertebrate aggressors, with specific interactions showing reduviid bugs, chrysopid lacewings, and vespid wasps avoiding the saddleback caterpillars rather than attacking them (Murphy et al. 2009).

During pupation, the silken cocoon is impregnated with calcium oxalate that is excreted from the maphigian tubules; this hardens the cocoon against disturbance (Ishii et al. 1984). Care should be taken if around a cocoon of *Acharia* sp., as this genus is documented to weave urticating spines into the silk layer as well (Epstein 1996; Scott 1963).

Medical Significance

Acharia stimulea is best known as a medically significant species. The large spines and potent hemolytic venom rank it as one of the most important North American species of urticating caterpillars, with larvae from the moth family Megalopygidae being the only lepidopterans considered more dangerous (Scott 1963, 1964; Durden and Mullen 2009; Hossler 2010).

The spines of *A. stimulea* are strong, acutely pointed, and hollow. They embed deeply into tissue and break off, and can interrupt healing as the protoplasm from the venom glands dries into the tissue area (Gilmer 1925). The venom itself can cause a systemic condition called erucism or acute urticaria, for which severe symptoms may include migraines, gastrointestinal symptoms, asthma complications, anaphylactic shock, rupturing of erythrocytes, and hemorrhaging (USAF 1982; Hossler 2009).

Physically manifested symptoms may or may not be present with erucism (Hossler 2009). Contact dermatitis caused by *A. stimulea* includes immediate intense burning sensations around the contact zone, arector pili muscles tightening causing hair to stand on end, increased perspiration in the affected area, red blanching of the skin, and blistering

(Edwards et al. 1986; Hossler 2009, 2010). Symptoms can last for five hours, and leave red blotches in the envenomation site (Hossler 2009).

Prevention of stings is the most effective measure when faced with erucism hazards: protect skin with long sleeves, pants, and gloves when anticipating potential contact with *A. stimulea*, and avoid areas of heavy infestation if known (USAF 1982; Hesler et al. 1999; Gottschling et al. 2007; Hossler 2009, 2010). Should contact occur, the removal of spines should always take precedence as the longer the spines are embedded in tissue the more venom is administered (Hossler 2009, 2010). Other symptoms should be treated on a case by case basis tailored for your specific reaction by a certified physician (Hossler 2009, 2010).

Personal repellants, such as DEET, are not effective controls for *A. stimulea*. Manual removal of eggs, larvae or cocoons with proper protective wear and tools is an effective method of control in case of unpredicted population increase (Benaim-Pinto et al. 1991; Gottschling et al. 2007; Iserhard et al. 2007). Spines of urticating caterpillars can become airborne and consequently be inhaled or contact sensitive tissues like the eyes and nose. They may also embed in surfaces such as wooden tables and plastics, which become a contact hazard at a later time if the area is not cleaned. Stray spines can also get caught in fabrics, such as carpet, aprons and clothing and come into skin contact that way (Scott 1963; USAF 1982; Hossler 2009, 2010). If handling urticating caterpillars regularly, such as in rearing facilities, ensure you have access to a frequently sanitized, well ventilated work environment to mitigate hazards of loose spines (Hossler 2009).

Pest Significance

Landscaping

The polyphagous nature of *A. stimulea* can cause it to be present in a large variety of ornamentals or backyard garden situations (see previous list). Despite this, it is not a significant defoliator of any one plant species (Cassani et al. 1989; Howard et al. 2001; Costello et al. 2003). *Achardia stimulea* is a venom hazard related to working around plants it may select for oviposition. Wearing appropriate skin covering and manual removal of offenders are options available to those working in the landscape or garden.

Agricultural

As above, the polyphagous nature of *A. stimulea* has restricted its significance as a defoliator of any one plant. It is interesting to note that *A. stimulea* is found on more

species of palms, including crop palms such as those which produce coconut or oils, than any other limacodid (Howard et al. 2001). Because of the adhesive abilities and strong physical and chemical defenses, *A. stimulea* is not likely to avoid disturbance or danger, which puts plant handlers at higher risk for contact. The lack of nutrition yielded by palms in particular would also encourage longer feeding intervals by the larvae, leading to higher incidence of contact (Howard et al. 2001). Pest control options such as *Bacillus thuringiensis* and diflubenzuron have been used to control massive infestations of urticating caterpillars with success (DeLong 1981; USAF 1982; Scholz et al. 1993; Dunlop and Freeman 1997; Hessler et al. 1999; Balit et al. 2001).

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