

Anthurium Diseases: Identification and Control in Commercial Greenhouse Operations¹

David J. Norman and Gul Shad Ali²

Introduction

Because of its attractive, long-lasting flowers, *Anthurium* is popular as both an exotic cut-flower crop and as a flowering potted-plant crop. Growers most often report two bacterial diseases and three fungal diseases in their commercial greenhouse environments. This article provides guidelines to identify and treat diseases that may be encountered during commercial greenhouse production of *Anthurium*.

Anthurium

Commonly known as flamingo flower, Hawaiian love plant, cresta de gallo, or tongue of fire, *Anthurium* has nearly 1,000 species, making it the largest genus in the plant family Araceae. *Anthurium* is native to tropical America, Mexico, Costa Rica, Cuba, and Brazil. Growth habits vary depending on species; some are terrestrial, others are epiphytic (Chen et al. 2003).

Most cut-flower *Anthurium* cultivars are selections of *Anthurium andraeanum*, an epiphytic-growing plant native to Columbia and Ecuador. The large red flowers produced by *Anthurium andraeanum* cultivars are very recognizable to consumers (Figure 1). Breeding has introduced such



Figure 1. Anthurium 'Kozohara' used in cut-flower production. Credits: D. Norman, UF/IFAS

flower colors as pink, orange, white, green, purple, and combinations of these colors.

Florida is now leading the nation in flowering potted *Anthurium* production. Cultivars for potted plant production have been derived from crosses of *A. andraeanum* with dwarf species, such as *A. amnicola* and *A. antioquiense*. Compact, hybrid varieties of potted *Anthurium* released by the University of Florida Plant Breeding Program include

- 1. This document is PP292, one of a series of the Plant Pathology Department, UF/IFAS Extension. Original publication date February 2012. Reviewed March 2015. Revised July 2018. Visit the EDIS website at http://edis.ifas.ufl.edu.
- 2. David J. Norman, associate professor; and Gul Shad Ali, assistant professor, Plant Pathology Department, Mid-Florida Resesarch and Education Center, UF/IFAS Extension, Gainesville, FL 32611.

The use of trade names in this publication is solely for the purpose of providing specific information. UF/IFAS does not guarantee or warranty the products named, and references to them in this publication do not signify our approval to the exclusion of other products of suitable composition. All chemicals should be used in accordance with directions on the manufacturer's label. Use pesticides safely. Read and follow directions on the manufacturer's label.

The Institute of Food and Agricultural Sciences (IFAS) is an Equal Opportunity Institution authorized to provide research, educational information and other services only to individuals and institutions that function with non-discrimination with respect to race, creed, color, religion, age, disability, sex, sexual orientation, marital status, national origin, political opinions or affiliations. For more information on obtaining other UF/IFAS Extension publications, contact your county's UF/IFAS Extension office.

U.S. Department of Agriculture, UF/IFAS Extension Service, University of Florida, IFAS, Florida A & M University Cooperative Extension Program, and Boards of County Commissioners Cooperating. Nick T. Place, dean for UF/IFAS Extension.

'Red Hot' (Henny, Chen, and Mellich 2008a), 'Orange Hot' (Figure 2) (Henny, Chen, and Mellich 2008b), and 'Southern Blush' (Henny, Poole, and Conover 1988).



Figure 2. *Anthurium* 'Orange Hot' used in flowering potted plant production.

Credits: R. J. Henny, UF/IFAS

Anthurium is very susceptible to bacterial and fungal diseases that can seriously limit commercial production. Bacterial blight caused by *Xanthomonas* is probably the most serious. Root rots caused by *Rhizoctonia*, *Pythium*, and *Phytophthora* also occur in *Anthurium* production. It is therefore important to be able to identify and eliminate these diseases.

Bacterial Diseases of *Anthurium* **Bacterial Blight**

Symptoms: The first visible symptoms are yellowed (chlorotic), water-soaked lesions along the leaf margins that grow rapidly to form dead (necrotic) V-shaped lesions characteristic of this disease (Figure 3).

The bacteria infect *Anthurium* plants by entering pores (hydathodes) along the leaf margins (Figure 4). Bacteria may also enter if leaf tissues become torn through pruning, or if leaf tissues are punctured by insects. When flowers are harvested, bacteria can enter via wounds.

Guttation droplets form at night when humidity is high and potting soil is warm and wet. Amino acids found in this guttation fluid are a source of food for the invading bacteria. Some infected plants are asymptomatic (do not show any disease symptoms) for months as the bacteria multiply. Bacteria in the guttation fluid from these asymptomatic infected plants can infect adjacent plants.



Figure 3. *Xanthomonas* bacterial blight exhibits characteristic V-shaped, water-soaked lesions forming along the edges on *Anthurium* leaves.

Credits: D. Norman, UF/IFAS



Figure 4. *Xanthomonas* bacteria enter the leaf margins via pores (hydathodes) where guttation droplets form.

Credits: D. Norman, UF/IFAS

Invading bacteria quickly spread throughout the plant (Figure 5). Leaves of systemically infected plants may exhibit a bronzed appearance. Floral quality may be reduced and/or flowers may become unmarketable (Figure 6). Eventually, plants wilt and die.

Causal Agent: Xanthomonas axonopodis pv. dieffenbachiae

The species of *Xanthomonas* that infects *Anthurium* has a very broad host range and is able to infect most aroid species; therefore, *Anthurium* plants may get blight when grown in close proximity to other aroids, such as *Dieffenbachia*, *Aglaonema*, and *Spathiphyllum*.



Figure 5. Extensive necrosis develops on *Anthurium* plants that are systemically infected with *Xanthomonas* bacterial blight. Credits: D. Norman, UF/IFAS



Figure 6. Xanthomonas bacterial blight lesions can also appear on the flowers.

Credits: D. Norman, UF/IFAS

Factors Favoring the Disease: Bacteria can swim across wet surfaces; therefore, it is very important to keep the foliage dry. The most effective way of accomplishing this is through drip irrigation.

Control and Treatment: Lower greenhouse humidity and temperature by increasing air circulation and venting the production facility. Allow space between the plants on the bench. Warm temperatures, high humidity, and saturated soils contribute to the formation of guttation droplets.

Only clean, tissue-cultured plantlets should be used when establishing new plantings. Because *Xanthomonas* bacteria can enter the plant through any wound or tear in the stem or foliage, the disease is easily spread if propagation is done via cuttings or division.

The disease can be spread when harvesting flowers or removing old foliage. Sterilize knives and clippers by dipping cutting tools in a disinfectant between plants. The most effective disinfectants are the quaternary ammonium compounds. It is also good practice to have two knives or shears in a dip bucket so they can be alternated, thus extending time in disinfectant and allowing for better coverage of the cutting surface.

When infected plants are found, they should be discarded immediately.

Products containing copper (Kocide® 3000, Phyton 27®, Camelot), mancozeb (Protect T/O^{TM} , Dithane®), and *Bacillus subtilis* (Cease®) are effective against *Xanthomonas*.

Bacterial Wilt

Symptoms: Leaf yellowing (chlorosis) is usually the first symptom observed. The disease spreads rapidly throughout the vascular system of the plant, turning veins in the leaves and stems a brown, bronze color (Figure 8). Bacterial ooze (brown slime) will be present if cuts are made into the stems of highly infected plants. Plants will exhibit wilt symptoms even though adequate soil moisture is available.



Figure 7. *Ralstonia* bacterial wilt causes yellowing (chlorosis) of *Anthurium* leaves.

Credits: D. Norman, UF/IFAS

Causal Agent: Ralstonia solanacearum

Ralstonia is known to infect several hundred plant genera. This bacterial disease is an opportunistic pathogen that colonizes soil and can remain viable for years without a host plant.

Factors Favoring the Disease: Cool greenhouse temperatures may temporarily mask symptoms and give bacteria time to spread. Symptoms appear rapidly during hot weather.



Figure 8. Wilt symptoms are the byproduct of *Ralstonia* bacteria clogging the vascular system of the plant.

Credits: D. Norman, UF/IFAS

Control and Treatment: A strict sanitation program is the most successful way to stop the spread of this pathogen and eventually eradicate it from a production facility. Fungicides that contain phosphorous acid have also been shown to be effective in preventing infection; however, they do not cure systemically infected plants (Norman et al. 2006).

Bacterial wilt is spread via contaminated soil, water, tools, or worker contact. Use disease-free propagation material. The bacterial wilt pathogen is easily spread via infected cuttings. Because the bacteria survive well in soil, both contaminated plant material and the supporting soil should be discarded. If pots and trays from contaminated infected plants are to be reused, they should be scrubbed free of adhering soil and then soaked in a disinfectant to kill the remaining bacteria. Knives and clippers should be sterilized between plants with a disinfectant containing a quaternary ammonium compound (Physan 20™, Green-Shield®) or diluted solution of bleach to prohibit spread.

Fungal Diseases of Anthurium Rhizoctonia Root Rot

Symptoms: The term "damping-off" is used to describe these classical symptoms. Young, tender stems are girdled, become water soaked, and are unable to support the weight of the plant (Figure 9). *Rhizoctonia* attacks the roots and lower stems of plants (Figure 10), but under wet conditions it can also attack and spread in the upper leaf canopy.

Causal Agent: Rhizoctonia solani

Rhizoctonia can survive within soil for years without a host plant. The fungus produces small mats of tightly woven mycelia called sclerotia. Sclerotia are irregular in shape, brown in color, and resemble particles of soil. Sclerotia



Figure 9. Anthurium wilt caused by Rhizoctonia root rot. Credits: D. Norman, UF/IFAS



Figure 10. Discolored brown roots are one of the symptoms observed with *Rhizoctonia* infections.

Credits: D. Norman, UF/IFAS

provide a seedlike mechanism for the fungus to survive unfavorable conditions, such as drought or cold weather. These small sclerotia stick to trays and pot surfaces and are one of the ways the fungus spreads through nurseries.

Factors Favoring the Disease: Water-saturated soils are conducive to disease development.

Control and Treatment: Never incorporate native soils into media mixes without steam sterilizing. Use well-drained soil mixes. Never store peat moss, sphagnum moss, chips, or potting media mixes directly on soil surfaces where they can be colonized by the fungus. Plants should be cultivated on raised benches to limit root contact with soil. *Rhizoctonia* frequently gains access to production facilities via infected propagation material.

Many fungicides are effective against outbreaks of *Rhizoctonia*. Examples include 3336®, OHP 6672® (thiophanate methyl), Medallion® (fludioxonil), and Prostar® (flutolanil).

Phytophthora/Pythium

Both *Phytophthora* and *Pythium* are called "oomycetes," commonly known as water molds. The control, spread, and management recommendations for *Phytophthora* and *Pythium* infestations are the same. Plants infected with either of these pathogens exhibit wilting, leaf yellowing (chlorosis), and root dieback (Figure 11).



Figure 11. *Anthurium* wilt caused by *Phytophthora*. Credits: D. Norman, UF/IFAS

Symptoms: *Phytophthora* and *Pythium* infections primarily attack root systems. Plants will wilt even though adequate soil moisture is available. Root sloughing is the primary diagnostic tool (Figure 12). Under severe conditions, the foliage may exhibit black to brown leaf lesions.

These symptoms are similar to symptoms caused by *Rhizoctonia*; however, fungal strands (mycelial growth) are rarely observed with *Phytophthora* or *Pythium* infections.

Causal Agents: *Phytophthora nicotianae* var. *parasitica* and *Pythium splendens*

Phytophthora and *Pythium* species cause substantial damage to *Anthurium* and numerous tropical foliage crops.

Factors Favoring the Disease: Water-saturated soils are conducive to disease development. These diseases can usually be avoided by using light, well-drained soil mixes.



Figure 12. *Pythium* and *Phytophthora* cause root sloughing. Credits: D. Norman, UF/IFAS

Control and Treatment: Use well-drained, synthetic soil mixes. Use disease-free stock plants. Plants with symptoms of disease should be discarded and the rest of the production facility should be treated with a fungicide drench. If potting containers are reused, they should be scrubbed and sterilized. Cutting shears, knives, and tools should be dipped in an appropriate disinfectant between plants.

Fungicides such as mefenoxam (Subdue® Maxx®) and aluminum tris/Fosetyl-al (Aliette® WDG), dimethomorph (Stature®), fluopicolide (Adorn $^{\text{\tiny M}}$), and phosphorous acid (Alude $^{\text{\tiny M}}$, K-Phite®) may be used to control *Phytophthora* and *Pythium*.

Black Nose Disease

Symptoms: Black nose can cause havoc in cut-flower and potted-plant production. Flowers and flowering potted plants cannot be sold with this condition.



Figure 13a. Black nose disease on *Anthurium* causes spadix darkening. Credits: D. Norman, UF/IFAS

The first symptoms observed are small, brown to black flecks on the floral spadix (nose) (Figure 13). These spots rapidly enlarge, become watery, turn brown to black, and may totally encompass the spadix. The spadix may eventually fall off. Growers may also observe black, sporecontaining structures (acervuli) on dead leaves and stems.



Figure 13b. Credits: D. Norman, UF/IFAS

Causal Agent: Colletotrichum gloeosporioides

The fungus *Colletotrichum gloeosporioides* attacks many temperate and tropical crops and can cause damage to roots, stems, leaves, and flowers. However, in *Anthurium* the pathogen is highly specific, only attacking the spadix portion of the flower (the nose).

Factors Favoring the Disease: This disease is most severe during humid, warm conditions. *Colletotrichum* readily invades plant tissues previously damaged by pesticides, fertilizer, or bacterial blight (*Xanthomonas*).

Control and Treatment: The *Colletotrichum* fungus produces thousands of small hot-dog-shaped spores that can readily be moved by splashing water, air movement, and workers. A strict sanitation program is crucial to control the spread of this pathogen in a production facility.

Fungicides containing mancozeb (Protect T/O^{TM} , Dithane®) are effective. Fungicide applications are usually discouraged because chemical residues diminish the marketability of flowers and plants.

Anthurium plant breeding programs both in Hawaii and Florida have incorporated disease resistance into many of

the current cultivars. Newer cultivars are highly resistant to this pathogen and rarely exhibit black nose.

References

Chen, J., D. B. McConnell, R. J. Henny, and K. C. Everitt. 2003. *Cultural Guidelines for Commercial Production of Interiorscape* Anthurium. ENH956. Gainesville: University of Florida Institute of Food and Agricultural Sciences. http://edis.ifas.ufl.edu/EP159

Henny, R. J., J. Chen, and T. A. Mellich. 2008a. *New Florida Foliage Plant Cultivar: 'Red Hot'* Anthurium. ENH1009. Gainesville: University of Florida Institute of Food and Agricultural Sciences. http://edis.ifas.ufl.edu/EP363

Henny, R. J., J. Chen, and T. A. Mellich. 2008b. *New Florida Foliage Plant Cultivar*: 'Orange Hot' Anthurium. ENH1100. Gainesville: University of Florida Institute of Food and Agricultural Sciences. http://edis.ifas.ufl.edu/EP364

Henny, R. J., R. T. Poole, and C.A. Conover. 1988. "Southern Blush' Anthurium." HortScience 23(5): 922–923.

Norman, D. J., J. Chen, A. Mangravita-Nova, and J. M. F. Yuen. 2006. "Control of Bacterial Wilt of Geranium with Phosphorus Acid." *Plant Dis.* 90(6): 798–802.