

Contaminants in the Urban Environment: Pharmaceuticals and Personal Care Products (PPCPs)—Part 2¹

Yun-Ya Yang, Gurpal S. Toor, and Alexander J. Reisinger²

This publication is part of a series titled Contaminants in the Urban Environment. This series is intended to give State and local government officials, soil scientists, consulting engineers, Extension agents, and citizens (1) a basic understanding of the occurrence, toxic effects, and source of various contaminants in the environment and (2) guidance on ways to protect human and environmental health.

This publication is part of a series titled Contaminants in the Urban Environment. This series is intended to give State and local government officials, soil scientists, consulting engineers, Extension agents, and citizens (1) a basic understanding of the occurrence, toxic effects, and source of various contaminants in the environment and (2) guidance on ways to protect human and environmental health.

Introduction and Purpose

Pharmaceuticals are products that we ingest or give to our pets and domesticated animals to improve health and to prevent or treat human and animal diseases. Pharmaceuticals include over-the-counter drugs, prescription medicines, nutritional supplements, and veterinary drugs.

Personal care products are products used to improve the quality of daily life by adorning and cleaning our bodies. Personal care products include lotions, fragrances, shampoos, antibacterial soaps, detergents, sunscreens, insect repellents, and cosmetics.

Together, these products are known as *pharmaceuticals* and *personal care products*, abbreviated as PPCPs (Figure 1).





Figure 1. Common pharmaceutical and personal care products (PPCPs) in households.

Credit: iStock/Thinkstock.com

Contaminants in the Urban Environment: Pharmaceuticals and Personal Care Products (PPCPs)—Part 1 provided an overview of PPCPs use and sale in the United States and the world (https://edis.ifas.ufl.edu/ss632). This publication, Contaminants in the Urban Environment: Pharmaceuticals and Personal Care Products (PPCPs)—Part 2, discusses the sources and impacts of PPCPs as well as the ways to protect our environment from PPCPs.

Sources of PPCPs in the Environment

PPCPs are released to the environment by multiple sources (Figure 2). Human contribution of PPCPs includes excretion in urine and feces and washing in baths and sinks inside the home. Other sources of PPCPs in the environment include residues from pharmaceutical manufacturing plants (which are well defined and controlled), residues from hospitals, illicit drug use, veterinary drug use (especially antibiotics and steroids), and agricultural activities.

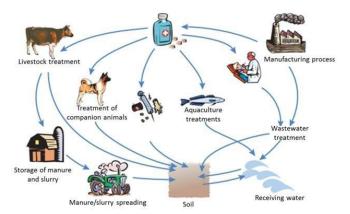


Figure 2. Routes of PPCPs release into the environment. Credit: Boxall (2004)

Among all these sources, human contribution of PPCPs is the major source. Human PPCPs are generally absorbed by the body after intake and are then excreted and discharged into the sewage or septic systems (wastewater). When wastewater is either discharged directly into the environment (after undergoing proper treatment at a WWTP), used for irrigation (i.e., reclaimed water) or biosolids (i.e., properly treated sewage sludge) and then applied as a fertilizer to agricultural land, PPCPs can be released into the environment (Kinney et al. 2006).

Veterinary and agricultural pharmaceuticals can be released to the environment via animal excretion or during the land application of manure. The usage and subsequent release of pharmaceuticals into the environment from animal usage is particularly concerning as approximately 80% of all antiobiotic usage in the United States is for agriculture/aquaculture (Hollis and Ahmed 2013). Another potential but less recognized source of PPCPs in groundwater is septic or onsite wastewater treatment systems. Septic systems remain important sources of PPCP contamination of groundwater, particularly where groundwater tables are shallow, such as in Florida and in several coastal states. To learn more about chemical compounds in septic systems, consult the EDIS article, Onsite sewage treatment and disposal systems: Trace Organic Chemicals at: https://edis.ifas.ufl.edu/ss554.

Impact of PPCPs in the Environment

A number of studies have documented the presence of PPCPs such as antibiotic, anti-inflammatory, anti-epileptic, and recreational medicines in ground- and surface waters throughout the world (see summary of these studies in Tables 1 and 2). The six most commonly

reported PPCPs globally in the groundwater (Table 1) are as follows (Lapworth et al. 2012):

- Caffeine (14 studies, average maximum concentration: 9,800 nanograms per liter [ng/L])
- Carbamazepine (23 studies, average maximum concentration: 5,000 ng/L)
- Bisphenol A (9 studies, average maximum concentration: 2,500 ng/L)
- Ibuprofen (14 studies, average maximum concentration: 1,500 ng/L)
- Sulfamethoxazole (15 studies, average maximum concentration: 252 ng/L)
- Diclofenac (11 studies, average maximum concentration: 121 ng/L)

These six compounds are also present in surface water, stream water, and finished drinking water (Table 2). Concentrations of PPCPs detected in surface waters near discharges of sewage effluents are as follows (Whitacre 2011):

- Caffeine (8 studies, minimum concentrations: 4,900 ng/L)
- Carbamazepine (9 studies, minimum–maximum concentrations: <1,000 to 7,100 ng/L)
- Ibuprofen (18 studies, minimum–maximum concentrations: non-detected (ND) to 1,200 ng/L)
- Sulfamethoxazole (10 studies, minimum-maximum concentrations: ND to 1,900 ng/L)
- Diclofenac (14 studies, minimum–maximum concentrations: ND to 5,040 ng/L)

Table 1 shows the frequency and summary statistics of PPCPs detected in regional and national scale groundwater reconnaissance studies. In these studies, the range of detection frequency was variable: sulfamethoxazole (10%–24%), ibuprofen (0%–7%), carbamazepine (1.2%-42%), bisphenol-A (8%-59%), and caffeine (13%-83%). The widespread occurrence of PPCPs in the environment could be due to their history of use and because they have had sufficient time to travel through the unsaturated soil zone (vadose zone) to the groundwater. In addition, properties of PPCPs can control the attenuation of PPCPs during wastewater treatment and the subsequent release of PPCPs into the environment. Even if PPCPs are partially or fully degraded in the environment or during treatment processes, the continual release of these contaminants

into the environment has led to their being termed *pseudopersistant* (Ellis 2006).

A summary of PPCPs' occurrence and frequency in the United States waters is provided in Table 2. The main classes of PPCPs in US freshwaters were antibiotic, antiinflammatory, anticonvulsant, sterol, flame retardant, plasticizer, and stimulant. Less information on PPCPs in the environment was found in the literature a few decades ago due to lack of knowledge and the analytical challenges of detecting PPCPs at trace levels. Note that these compounds might have existed for a long time since some of them (e.g., carbamazepine, sulfamethoxazole, and ibuprofen) have been approved for use in the United States for more than three decades. Due to the improvement in analytical methods in the last 15+ years, researchers now have the ability to measure very small concentrations at ng/L (parts per trillion).

Although environmental concentrations of PPCPs are low, the risks to human health and the environment cannot be excluded because they are still poorly understood. Research suggests that certain PPCPs (e.g., hormones) may cause reproductive and developmental harm in aquatic organisms even at trace levels (ng/L to sub- μ g/L); these are the levels that are frequently observed in ground- and surface waters (Vajda et al. 2008). Furthermore, a wide range of studies have found that environmentally realistic concentrations of a variety of PPCPs can have sub-lethal ecological impacts (reviewed by Richmond et al. 2017).

Impacts on Aquatic Health: Most of the research on the effects of PPCPs involves antibiotics and steroid hormones. Even though these chemicals are present only at trace levels (part per trillion, or ng/L), PPCPs can cause changes in the behavior and biochemistry (i.e., functioning) in aquatic animals such as fish, frogs, and mussels. More notably, estrogens (or hormones)—such as estrone, estradiol, and ethinyl estradiol—have been observed to cause feminization of male fish at concentrations as low as a few ng/L (Jobling et al. 2003; Vajda et al. 2008). For example, exposure to 2-10 ng/L of 17α-ethinylestradiol (primarily used in hormonal contraceptives) induced synthesis of vitellogenin (an egg yolk protein precursor found only in females) in male zebrafish (Danio rerio), Japanese medaka (Oryzias latipes), and Chinese rare minnow (Gobiocypris rarus) (Ma et al. 2005; Xu et al. 2008; Zha et al. 2007). In a study

of PPCP impacts on microbial communities, Rosi et al. (2018) found that cimetidine (an antacid and antihistamine) and ciprofloxacin (an antibiotic) both reduce microbial respiration in streams rarely exposed to these compounds, and that microbial community composition was drastically different for microbial communities exposed to ciprofloxacin.

In personal care products, triclosan is an antibacterial and antifungal agent present in products such as soaps, deodorants, and toothpastes. Triclosan and its degradation product, triclocarban, have been investigated mainly because of their relative high toxicity to aquatic organisms (Dussault et al. 2008; Ishibashi et al. 2004; Tamura et al. 2013). Triclosan and triclocarban were found to be toxic with the 50% effect concentration (dose at which 50% of the test population is killed) values from 4 to 35 $\mu g/L$ (72-hour exposure) for green algae, 7 to 230 $\mu g/L$ (48-hour exposure) for daphnia, and 45 to 340 $\mu g/L$ (96-hour exposure) for fish (Tamura et al. 2013). The marketing and development of triclosan-containing products was banned by the United States Food and Drug Administration in 2017.

Impacts on Human Health: Studies have shown that some PPCPs originating from human therapies are not completely eliminated in the sewage treatment systems and are discharged as contaminants into the receiving waters (Stackelberg et al. 2004; Ternes et al. 2002). There may be a potential risk of drinking water contamination by PPCPs, especially in urban areas with high sewage wastewater outputs and low surface-water flows.

Eighty percent of the antibiotics sold in the United States are used for animal agriculture (Hollis and Ahmed 2013). According to the Food and Drug Administration (FDA), farm animals ingest nearly 30 million pounds of antibiotics annually. Research has verified the occurrence of veterinary antibiotics in manure, agricultural fields, and surface water bodies (Kim et al. 2010; Pinheiro et al. 2013). Once PPCPs enter the soil, compounds with strong fixation and resistance to degradation remain in surface soils for a long time. These compounds then could be potentially taken up by plants (Holling et al. 2012; Sabourin et al. 2012) and leach or run off to ground- and surface waters.

A relation between use of antibiotics and the occurrence of antibiotic-resistance in bacterial isolates from manure or manure-contaminated surface water has been found (Heuer and Smalla 2007; Peak et al. 2007). Antibiotic-resistant bacteria—that is, the bacteria that cannot be controlled or killed by antibiotics—can cause serious disease and are an important public health issue. Antibiotic-resistant bacteria attach to crops or become endophytes (i.e., bacteria living within plants) that may lead to exposure to humans through food supply. For more information on antibiotic resistance and food safety, see the Centers for Disease Control and Prevention website (2014;

http://www.cdc.gov/narms/faq.html).

Some chemicals and bacteria in food may represent hazards but at trace levels do not represent a human health risk, and research has yet to demonstrate an impact on human health from the trace levels at which PPCPs occur in the environment. However, according to the National Institute of Environmental Health Sciences (NIEHS), endocrine disruptors may pose the greatest risk during prenatal and early development when organ and neural systems are forming.

What do we Not know about PPCPs?

To date, the improved methods and progress in technology has made it possible to detect very low levels of PPCPs in the environment and their effects on aquatic and terrestrial organisms. However, there are still many questions that need to be addressed before we can eventually determine whether residues of PPCPs in the environment are a threat to human and environmental health.

First, how much of an impact do PPCPs have in the natural environment? For example, a related issue is whether we should worry about transformation products. There is sparse information on the formation of PPCP degradation products, the ecotoxicity of PPCP metabolites, and mixtures of PPCPs. In some instances, these degradation products may be more ecotoxic, more persistent, and more mobile than the parent compounds. Furthermore, many of the environmental impacts of PPCPs are sublethal. Although these sublethal effects may not directly kill organisms, changes in organismal behavior or function can have subsequent environmental impacts (Richmond et al. 2017). During risk assessment, an improved understanding of the sublethal effects and transformation of PPCPs would help identify the circumstances in which metabolites should receive closer attention.

Second, does environmental exposure to PPCP residues result in more antibacterial resistance? To help determine how important PPCP residues are in relation to public health, we would need a better understanding of (1) the role of environmental antibiotic residues in the selection of antibiotic resistance and (2) the potential for antibiotic resistances to transfer from the environment to human and animal pathogens.

Finally, what wastewater treatment methods are effective in reducing the effects of PPCPs in the environment? It may be necessary to develop advanced treatment systems that consider local conditions and constraints in order to remove PPCPs during the treatment processes. If the wastewater treatment methods were dramatically improved, then most of the PPCPs would never enter the environment. We suggest that this process should be a priority in our ongoing efforts to manage PPCPs, although this would require considerable amount of resources and infrastructure and the willingness of elected officials.

What can you do to protect the environment?

We all can use common-sense approaches to protect the environment. For example, do not flush unused or expired drugs down the drain. Proper disposal of drugs is a straightforward way for us to prevent pollution. The United States Environmental Protection Agency offers guidelines on proper disposal procedures for PPCPs, which can be access here:

https://www.epa.gov/sites/production/files/2015-06/documents/how-to-dispose-medicines.pdf. Some recommendations for consumers (US EPA 2009) include the following:

- · Keep drugs in the original container.
- · Scratch out patient information on labels.
- Place liquids in glass bottles and then in plastic resealable bags to contain leakage.
- Add some water to dry tablets or capsules to dissolve them slightly.
- Place the items in the trash.
- Check to see if your pharmacy has a drug "take back" program.
- Try to purchase only the quantity of drugs that you will use
- Cut down on the amount of PPCPs that you use.

 Follow any specific disposal instructions associated with your prescription.

For more information about proper disposal of prescription drugs, consult this website: http://www.fda.gov/ForConsumers/ConsumerUpdates/ucm101653.htm.

Other potential approaches to reduce the amount of PPCPs released to the environment include the following:

- Introduction of improved wastewater treatment options.
- Medicine labels that provide information on possible environmental impact, allowing consumers to consider environmental outcomes if the medication is not disposed of correctly.
- Learning more about these issues and sharing the message.

Appendix: Calculating the Amount of PPCPs in Wastewater

We provide this tentative calculation as a way to determine the levels of compounds in the environment, if the occurrence data are not widely available.

The amount of PPCPs in wastewater varies by the type of compound and by time of the year. The load of a pharmaceutical in wastewater can be estimated with a model based on the sale volume, wastewater production, disposal rate of unused medicines, excretion rate, and removal efficiency by the wastewater treatment plant (Zhang and Geißen 2010). For example, carbamazepine is an anticonvulsant and mood-stabilizing medicine used primarily in the treatment of seizure disorders and neuropathic pain. It is one of the most frequently detected pharmaceuticals in water bodies and is known to be persistent in the environment with removal efficiencies less than 20% (Oosterhuis, Sacher, and ter Laak 2013).

The example below demonstrates the amount of carbamazepine that may be excreted using the sale, wastewater production, excretion, and removal efficiency data from the literature.

Predicted PPCP concentration in the wastewater effluent $(\frac{\text{nanograms (ng)}}{\text{liter (L)}})$

= Human excretion of PPCP in wastewater (Equation 1)+ Disposal of PPCP by flushing in toilet or sinks (Equation 2)

Equation 1.

Calculation of the human excretion of PPCPs, using Equation 1:

$$\begin{split} & Human\ excretion\ of\ PPCP\ (\frac{ng}{L}) \\ & = \frac{PPCP\ sale\ (\frac{kg}{year})\times (\frac{used\ PPCP}{PPCP\ sale})\times excretion\ rate}{Annual\ wastewater\ production(km^3)} \\ & \times\ (1-overall\ removal\ efficiency) \end{split}$$

Example 1a.

For example, sale volume of carbamazepine in 2007 was 124,556 kg in the United States. The annual wastewater production in 2007 was 61 km³ (Zhang and Geißen 2010). The ratio of used carbamazepine to the carbamazepine sale is assumed to be 0.67, as indicated in a survey (Bound and Voulvoulis 2005). Assuming that approximately 30% of carbamazepine is excreted in the original form (Lienert, Güdel, and Escher 2007; Zhang, Geißen, and Gal 2008); and thus the excretion rate of carbamazepine is set at 0.3. The overall removal rate of carbamazepine in sewage systems is set at 0.1 based on literature (10%–13% removal).

Substitution of these values in Equation 1:

Human excretion of PPCP
$$= \frac{124556(kg) \times 0.67 \times 0.3}{61(km^3)} \times (1 - 0.1)$$
$$= 369(\frac{kg}{km^3}) = 369(\frac{ng}{L})$$

Example 1b.

As 1 kg = 10^{12} ng and 1 km³ = 10^{12} L, this equation evaluates to 369 ng/L of carbamazepine in wastewater.

Equation 2

Calculation of the amount of PPCP that is directly disposed or flushed, using Equation 2:

Disposal of unused PPCP by flushing in toilet or sinks $(\frac{ng}{I})$

$$= \frac{PPCP \ sale \ (\frac{kg}{year}) \times (\frac{disposed \ PPCP}{PPCP \ sale})}{Annual \ wastewater \ production(km^3)} \times (1$$

$$- \ overall \ removal \ efficiency)$$

Equation 2.

In this case, the ratio of disposing carbamazepine to the carbamazepine sale is set at 0.17, as indicated in a survey on disposal practices of unused medications (Bound and Voulvoulis 2005). All other parameters will remain the same as human excretion of PPCP (Equation 1).

Disposal of PPCP by flushing in toilet or sinks

$$= \frac{124556 \left(\frac{kg}{year}\right) \times (0.17)}{61 \left(\frac{km^3}{year}\right)} \times (1 - 0.1)$$
$$= 312 \left(\frac{kg}{km^3}\right) = 312 \left(\frac{ng}{L}\right)$$

Example 2a.

Thus, the predicted concentration of carbamazepine in the sewage effluent/wastewater will be the sum of human excretion (369 ng/L), as calculated by Equation 1, and direct disposal (312 ng/L), as calculated by Equation 2, resulting in total concentration of 681 ng/L of wastewater.

The concentrations of other PPCPs in the wastewater can be calculated using the above equations, if other data used in equations 1 and 2 are available. This may provide a way to calculate the likely concentrations of other PPCPs in the environment.

References

Barnes, K. K., D. W. Kolpin, E. T. Furlong, S. D. Zaugg,
M. T. Meyer, and L. B. Barber. 2008. "A National
Reconnaissance of Pharmaceuticals and Other
Organic Wastewater Contaminants in the United
States - I) Groundwater." *Science of the Total*Environment 402: 192–200.

- Benotti, M. J., R. A. Trenholm, B. J. Vanderford, J. C.
 Holady, B. D. Stanford, and S. A. Snyder. 2009.
 "Pharmaceuticals and Endocrine Disrupting
 Compounds in US Drinking Water."

 Environmental Science and Technology 43: 597–603.
- Bound, J. P., and N. Voulvoulis. 2005. "Household Disposal of Pharmaceuticals as a Pathway for Aquatic Contamination in the United Kingdom."

 Environmental Health Perspectives 113: 1705–1711.
- Boxall, A. B. A. 2004. "The Environmental Side Effects of Medication." *EMBO Rep 5*: 1110–1116.
- Center for Disease Control and Prevention. 2013.

 "Frequently Asked Questions about Antibiotic
 Resistance and Food Safety." Accessed February
 10, 2014. http://www.cdc.gov/narms/faq.html.
- Dussault, È. B., V. K., Balakrishnan, E. Sverko, K. R. Solomon, and P. K. Sibley. 2008. "Toxicity of Human Pharmaceuticals and Personal Care Products to Benthic Invertebrates."

 Environmental Toxicology and Chemistry 27: 425–432.
- Ellis, J. B. 2006. "Pharmaceutical and personal care products (PPCPs) in urban receiving waters." Environmental Pollution 144: 184–189.
- Focazio, M. J., D. W. Kolpin, K. K. Barnes, E. T. Furlong, M. T. Meyer, S. D. Zaugg, L. B. Barber, and M. E. Thurman. 2008. "A National Reconnaissance for Pharmaceuticals and Other Organic Wastewater Contaminants in the United States II)

 Untreated Drinking Water Sources." *Science of the Total Environment* 402: 201–216.
- Fram, M. S., and K. Belitz. 2011. "Occurrence and Concentrations of Pharmaceutical Compounds in Groundwater Used for Public Drinking-Water Supply in California." *Science of the Total Environment* 409: 3409–3417.
- Heuer, H., and K. Smalla. 2007. "Manure and Sulfadiazine Synergistically Increased Bacterial Antibiotic Resistance in Soil Over at Least Two Months." *Environmental Microbiology* 9: 657–666.

- Holling, C. S., J. L. Bailey, B. V. Heuvel, and C. A. Kinney. 2012. "Uptake of Human Pharmaceuticals and Personal Care Products by Cabbage (*Brassica campestris*) from Fortified and Biosolids-Amended Soils." *Journal of Environmental Monitoring* 14: 3029–3036.
- Hollis, A., and Z. Ahmed. 2013. "Preserving antibiotics, rationally." *New England Journal of Medicine* 369: 2474–2476.
- Ishibashi, H., N. Matsumura, M. Hirano, M. Matsuoka, H. Shiratsuchi, Y. Ishibashi, Y. Takao, and K. Arizono. 2004. "Effects of Triclosan on the Early Life Stages and Reproduction of Medaka *Oryzias latipes* and Induction of Hepatic Vitellogenin."

 Aquatic Toxicology 67: 167–179.
- Jobling, S., D. Casey, T. Rodgers-Gray, J. Oehlmann, U. Schulte-Oehlmann, S. Pawlowski, T. Baunbeck, A. P. Turner, and C. R. Tyler. 2003.

 "Comparative Responses of Molluscs and Fish to Environmental Estrogens and an Estrogenic Effluent." *Aquatic Toxicology* 65: 205–220.
- Kim, S. C., J. G. Davis, C. C. Truman, J. C. Ascough, and K. Carlson. 2010. "Simulated Rainfall Study for Transport of Veterinary Antibiotics—Mass Balance Analysis." *Journal of Hazardous Materials* 175: 836–843.
- Kinney, C. A., E. T. Furlong, S. L. Werner, and J. D. Cahill. 2006. "Presence and Distribution of Wastewater-Derived Pharmaceuticals in Soil Irrigated with Reclaimed Water." *Environmental Toxicology and Chemistry* 25: 317–326.
- Kolpin, D. W., E. T. Furlong, M. T. Meyer, E. M.
 Thurman, S. D. Zaugg, L. B. Barber, and H. T.
 Buxton. 2002. "Pharmaceuticals, Hormones, and
 Other Organic Wastewater Contaminants in U.S.
 Streams, 1999–2000: A National
 Reconnaissance." *Environmental Science and*Technology 36: 1202–1211.
- Lapworth, D. J., N. Baran, M. E. Stuart, and R. S. Ward. 2012. "Emerging Organic Contaminants in Groundwater: A Review of Sources, Fate and Occurrence." *Environmental Pollution* 163: 287–303.

- Lienert, J., K. Güdel, B. I. Escher. 2007. "Screening Method for Ecotoxicological Hazard Assessment of 42 Pharmaceuticals Considering Human Metabolism and Excretory Routes."

 Environmental Science and Technology 41: 4471–4478.
- Ma, T., X. Wan, Q. Huang, Z. Wang, and J. Liu. 2005.

 "Biomarker Responses and Reproductive
 Toxicity of the Effluent from a Chinese Large
 Sewage Treatment Plant in Japanese medaka
 (Oryzias latipes)." Chemosphere 59: 281–288.
- Miller, K. J., and J. Meek. 2006. "Helena Valley Ground Water: Pharmaceuticals, Personal Care Products, Endocrine Disruptors (PPCPs) and Microbial Indicators of Fecal Contamination." Montana Bureau of Mines and Geology, Open File Report 532.
- National Center for Health Statistics. 2013. "Health, United States, 2012: With Special Feature on Emergency Care." Hyattsville, MD.
- Oosterhuis, M., F. Sacher, and T. L. ter Laak. 2013.

 "Prediction of Concentration Levels of
 Metformin and Other High Consumption
 Pharmaceuticals in Wastewater and Regional
 Surface Water Based on Sales Data." Science of
 the Total Environment 442: 380–388.
- Peak, N., C. W. Knapp, R. K. Yang, M. M. Hanfelt, M. S. Smith, D. S. Aga, and D. W. Graham. 2007.

 "Abundance of Six Tetracycline Resistance Genes in Wastewater Lagoons at Cattle Feedlots with Different Antibiotic use Strategies."

 Environmental Microbiology 9: 143–151.
- Pinheiro, A., R. M. Rosa Albano, T. C. Alves, V.

 Kaufmann, and M. R. da Silva. 2013. "Veterinary
 Antibiotics and Hormones in Water from
 Application of Pig Slurry to Soil." *Agricultural Water Management* 129: 1–8.
- Richmond, E. K., M. R. Grace, J. J. Kelly, A. J. Reisinger, E. J. Rosi, and D. M. Walters. 2017.

 "Pharmaceuticals and personal care products (PPCPs) are ecological disrupting compounds (EcoDC)." *Elementa: Science of the Anthropocene* 5: 66.

- Rosi, E. J., H. A. Bechtold, D. Snow, M. Rojas, A. J. Reisinger, and J. J. Kelly. 2018. "Urban stream microbial communities show resistance to pharmaceutical exposure." *Ecosphere* 9: e02041.
- Sabourin, L., P. Duenk, S. Bonte-Gelok, M. Payne, D. R. Lapen, and E. Topp. 2012. "Uptake of Pharmaceuticals, Hormones and Parabens into Vegetables Grown in Soil Fertilized with Municipal Biosolids." *Science of the Total Environment* 431: 233–236.
- Stackelberg, P. E., E. T. Furlong, M. T. Meyer, S. D. Zaugg, A. K. Henderson, and D. B. Reissman. 2004. "Persistence of Pharmaceutical Compounds and Other Organic Wastewater Contaminants in A Conventional Drinking-Water Treatment Plant." Science of the Total Environment 329: 99–113.
- Tamura, I., K. Kagota, Y. Yasuda, S. Yoneda, J. Morita, N. Nakada, Y. Kameda, K. Kimura, N. Tatarazako, and H. Yamamoto. 2013.

 "Ecotoxicity and Screening Level Ecotoxicological Risk Assessment of Five Antimicrobial Agents: Triclosan, Triclocarban, Resorcinol, Phenoxyethanol and p-thymol."

 Journal of Applied Toxicology 33: 1222–1229.
- Ternes, T. A., M. Meisenheimer, D. McDowell, F. Sacher, H. Brauch Jr., B. Haist-Gulde, G. Preuss, U. Wilme, and N. Zulei-Seibert. 2002. "Removal of Pharmaceuticals during Drinking Water Treatment." *Environmental Science and Technology* 36: 3855–3863.
- Toor, G. S., M. Lusk, and T. Obreza. 2011. *Onsite Sewage Treatment and Disposal Systems: Trace Organic Chemicals*. Gainesville: University of Florida Institute of Food and Agricultural Sciences. http://www.edis.ifas.ufl.edu/ss554.
- United States Environmental Protection Agency (US EPA). 2009. How to Dispose of Medicines Properly.
- United States Food and Drug Administration. 2013.

 "How to Dispose of Unused Medicines."

 Accessed February 10, 2014.

 http://www.fda.gov/ForConsumers/ConsumerUpdates/ucm101653.htm.

- United States Pharmaceutical Statistics. 2013. "US
 Pharmaceuticals Sales—2012." Accessed
 December 2, 2013.
 http://www.drugs.com/stats/top100/2012/sales.
- Vajda, A. M., L. B. Barber, J. L. Gray, E. M. Lopez, J. D. Woodling, and D. O. Norris. 2008. "Reproductive Disruption in Fish Downstream from an Estrogenic Wastewater Effluent." *Environmental Science and Technology* 42: 3407–3414.
- Whitacre, D. M. 2011. "Reviews of Environmental Contamination and Toxicology." *Springer* 202: 79–84.
- Xu, H., J. Yang, Y. Wang, Q. Jiang, H. Chen, and H. Song. 2008. "Exposure to 17α-ethynylestradiol Impairs Reproductive Functions of Both Male and Female zebrafish (*Danio rerio*)." *Aquatic Toxicology* 88: 1–8.
- Zha, J., Z. Wang, N. Wang, and C. Ingersoll. 2007.

 "Histological Alternation and Vitellogenin
 Induction in Adult Rare Minnow (*Gobiocypris rarus*) after Exposure to Ethynylestradiol and
 Nonylphenol." *Chemosphere* 66: 488–495.
- Zhang, Y., and S.-U. Geißen. 2010. "Prediction of Carbamazepine in Sewage Treatment Plant Effluents and Its Implications for Control Strategies of Pharmaceutical Aquatic Contamination." *Chemosphere* 80: 1345–1352.
- Zhang, Y., S.-U. Geißen, and C. Gal. 2008.

 "Carbamazepine and Diclofenac: Removal in
 Wastewater Treatment Plants and Occurrence
 in Water Bodies." *Chemosphere* 73: 1151–1161.

Tables

Table 1. Occurrence of selected PPCPs in reconnaissance groundwater studies.

Country/region	Detection frequency (%)	Max conc. (ng/L)	Number of samples
Sulfamethoxazole (ar	ntibiotic)		
US	23.4	1110	47
Europe	24.2	38	164
Switzerland	18	48	100
France	18	18	147
Germany	10	410	105
Ibuprofen (anti-inflam	matory)		
US	2.1	3110	47
Europe	6.7	395	164
Switzerland	0	_	47
France	0.5	7	209
Germany	0	_	105
UK	0.3	290	2644
Carbamazepine (antic	convulsant)		
US	1.5	420	1231
Europe	42.1	390	164
Switzerland	19	45	47
France	42	167	218
Germany	12	900	105
UK	1.2	3600	2644
Bisphenol-A (plasticiz	rer)		
US	29.8	2550	47
Europe	39.6	2299	164
UK	8	9300	2644
Austria	59	930	110
Caffeine (stimulant)	1	1	
US	12.8	130	47
Europe	82.9	189	164
UK	27	4500	2644
Source: Adapted from	n Lapworth et al. (2012)	1	

Table 2. PPCP residues in waters in the United States

Type of study (sites)	Source	Chemicals	Use	Max conc. (ng/L)	Frequency (%)
Reconnaissance survey: groundwater (25), surface water (49)	Surface water Groundwater	cholesterol	sterol	Not applicable (NA)	59
		cotinine	nicotine metabolite		51
(Focazio et al. 2008)		1,7-dimethylxanthine	caffeine		27
		carbamazepine	metabolite		20
		bisphenol-A	anticonvulsant		20
		1,7-dimethylxanthine	plasticizer		16
		tri(2-chloroethyl) phosphate	caffeine metabolite		12
			fire retardant		
Reconnaissance survey (139)	Stream water	acetaminophen	analgesic	10000	24
(Kolpin et al. 2002)		sulfamethoxazole	antibiotic	1900	13
		caffeine	stimulant	6000	61
		codeine	opioid analgesic	19	7
		ibuprofen	anti- inflammatory	1000	10
		bisphenol-A	plasticizer	12000	41
		tri(2-chloroethyl) phosphate	fire retardant	540	58
		17ß-estradiol	hormone	200	11
		estrone	hormone	112	7
		progesterone	hormone	199	4
		testosterone		214	3
		cholesterol	sterol	150000	86
Comprehensive survey (18)	Finished	atenolol	beta-blocker	18	44
(Benotti et al. 2009)	drinking water	carbamazepine	anticonvulsant	18	44
(25.000)		fluoxetine	antidepressant	0.82	11
		gemfibrozil	antilipidemic	2.1	39
		meprobamate	antianxiety	42	78
		phenytoin	anticonvulsant	19	56
		sulfamethoxazole	antibiotic	3	22
		triclosan	antibacterial	1.2	6
		bisphenol-A	plasticizer	25	6

Type of study (sites)	Source	Chemicals	Use	Max conc. (ng/L)	Frequency (%)		
Groundwater survey (1231)	Groundwater	acetaminophen	analgesic	1890	0.32		
(Fram and Belitz 2011)		caffeine	stimulant	290	0.24		
		p-xanthine	caffeine metabolite	120	0.08		
		carbamazepine	anticonvulsant	420	1.5		
		codeine	opioid analgesic	214	0.16		
		sulfamethoxazole	antibiotic	170	0.41		
		trimethoprim	antibiotic	18	0.08		
Groundwater survey (38)	Groundwater	carbamazepine	anticonvulsant	420	NA		
(Miller and Meek 2006)		dilantin	anti-epileptic	22			
		diclofenac	anti- inflammatory	46			
Nation-wide survey (47)	Groundwater	sulfamethoxazole	antibiotic	NA	23		
(Barnes et al. 2008)		bisphenol-A	plasticizer		30		
		tri(2-chloroethyl) phosphate	fire retardant		30		
		4-octylphenol monoethoxylate	detergent metabolite		19		
Source: Adapted from Lapworth et al. (2012)							

The Institute of Food and Agricultural Sciences (IFAS) is an Equal Opportunity Institution authorized to provide research, educational information and other services only to individuals and institutions that function with non-discrimination with respect to race, creed, color, religion, age, disability, sex, sexual orientation, marital status, national origin, political opinions or affiliations. For more information on obtaining other UF/IFAS Extension publications, contact your county's UF/IFAS Extension office. U.S. Department of Agriculture, UF/IFAS Extension Service, University of Florida, IFAS, Florida A & M University Cooperative Extension Program, and Boards of County Commissioners Cooperating. Andra Johnson, dean for UF/IFAS Extension.

¹This document is SL420, one of a series of the Department of Soil, Water, and Ecosystem Sciences, UF/IFAS Extension. Original publication date December 2014. Revised May 2018 and August 2021. Visit the EDIS website at https://edis.ifas.ufl.edu.

² Yun-Ya Yang, former postdoctoral researcher, Soil and Water Quality Laboratory, UF/IFAS Gulf Coast Research and Education Center, Wimauma, FL; Gurpal Toor, former associate professor, Department of Soil and Water Sciences, UF/IFAS Gulf Coast Research and Education Center, Wimauma FL 33598; Alexander J. Reisinger, associate professor of urban soil and water quality, UF/IFAS Department of Soil, Water, and Ecosystem Sciences, and state Extension specialist, Center for Land Use Efficiency, Program for Resource Efficient Communities; UF/IFAS Extension, Gainesville, FL 32611.